Single cell integration

Nathalie Lehmann







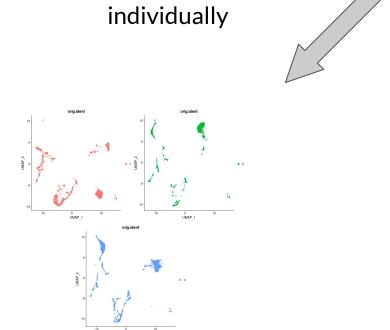
So far: worked on 1 individual matrix

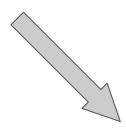
Generally: more than 1 sample

So far: worked on 1 individual matrix

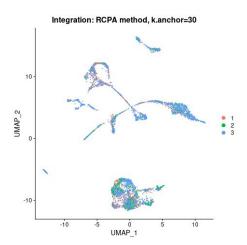
Generally: more than 1 sample

But should we study them





all samples together?

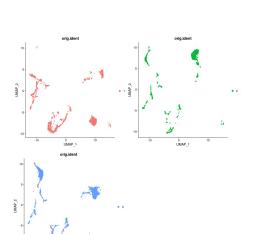


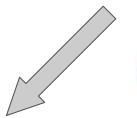
So far: worked on 1 individual matrix

Generally: more than 1 sample

But should we study them

individually







 Quick way to have a first look at data



- Repetitive
- Makes more sense to bring replicates together.
- Makes more sense to bring together similar samples (same experiment, organ...)

So far: worked on 1 individual matrix

Generally: more than 1 sample

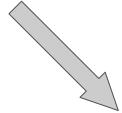
But should we study them



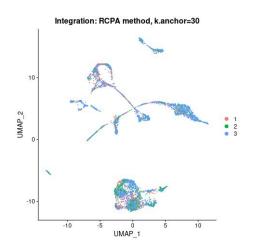
- Allows to work across multiple samples.
- Particularly important for cell populations visualization and identification
- Many cells: helps identifying rare populations



Overcorrection?



all samples together?

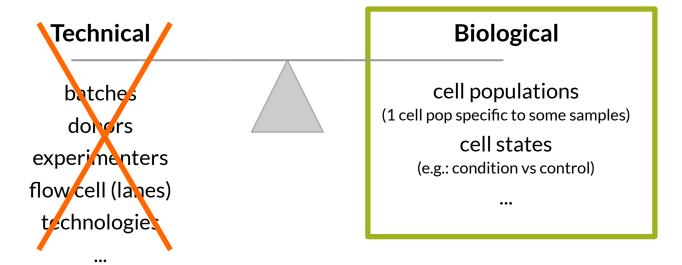


Question!

What do you think are the challenges in data integration?

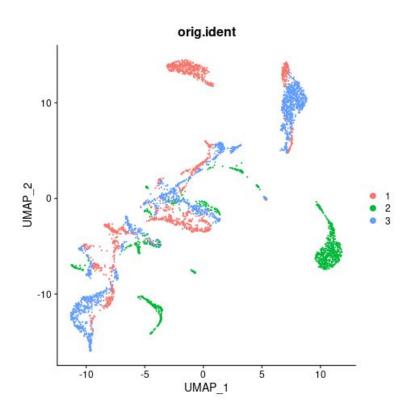
The challenge of data integration

2 sources of variability across samples



Question!

What do you think of this integration?



Question!

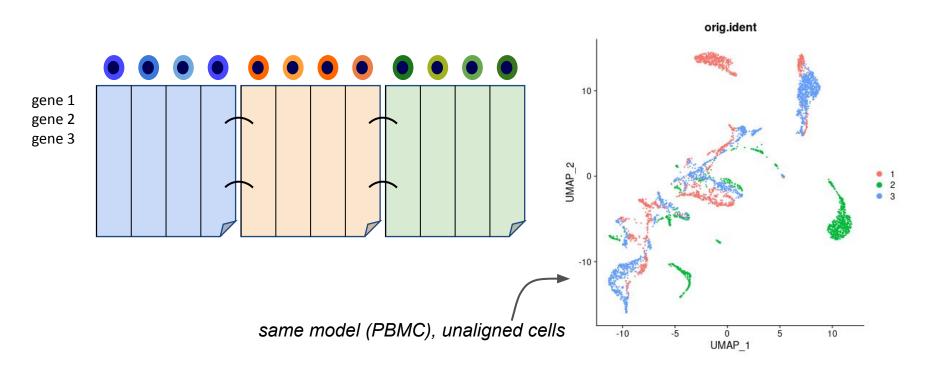
What do you think of this integration?



https://www.10xgenomics.com

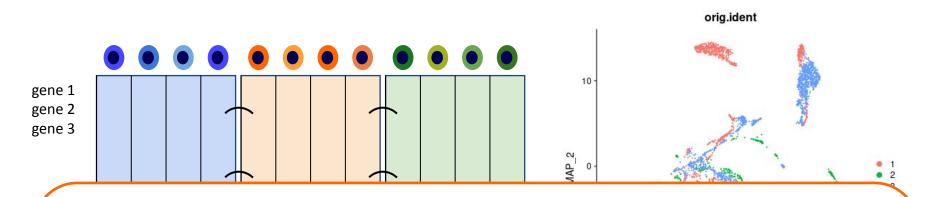
Problem...

...simple matrix concatenation does not always work



Problem...

...simple matrix concatenation does not always work



This is typically a problem of batch effect

We need a more sophisticated integration method

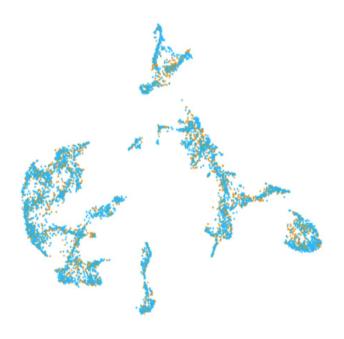
When to integrate

• Do not integrate

e.g.: replicates generated in the same time and exactly in the same manner may not need integration

PBMCs

sample 1 sample 2



https://www.10xgenomics.com

When to integrate?

Integrate

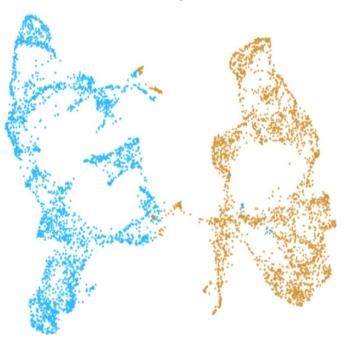
when obvious batch effect between samples, typically seen on low dimension visualization

In this example, the sample of origin would be a huge bias for clustering

The samples need integration to align cell types/clusters and then identify them correctly

PBMCs

sample 1 sample 2

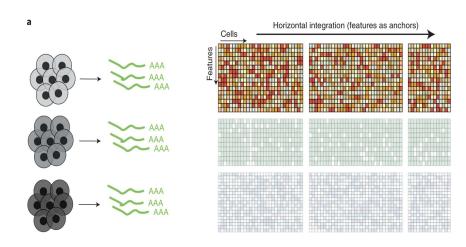


https://www.10xgenomics.com

Many methods

Different types of integrations

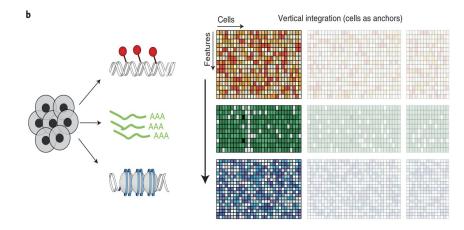
 Horizontal: different samples same modality



Different types of integrations

 Horizontal: different samples same modality

 Vertical: same sample different modalities (multiomics)

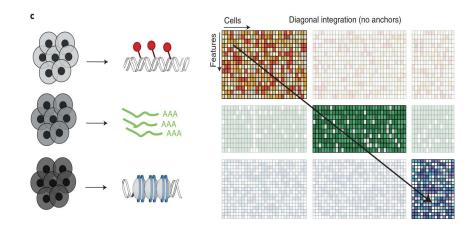


Different types of integrations

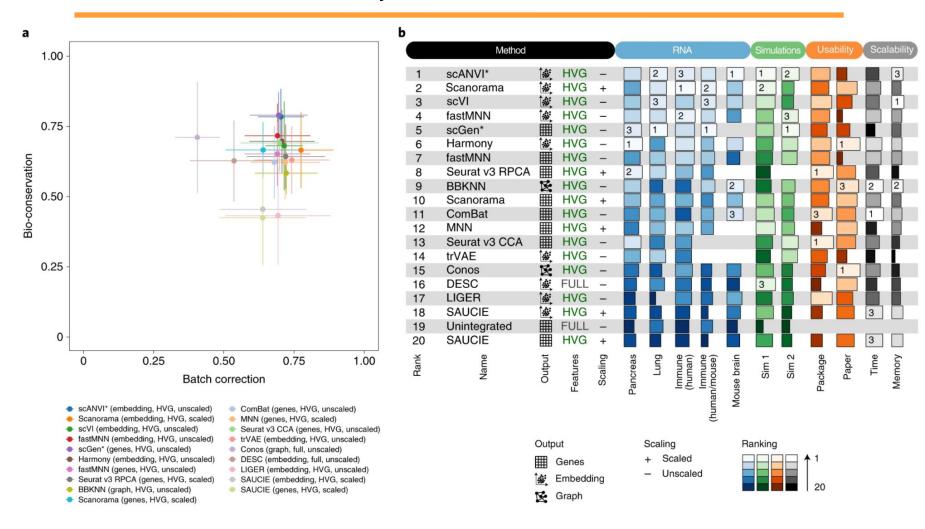
 Horizontal: different samples same modality

 Vertical: same sample different modalities (multiomics)

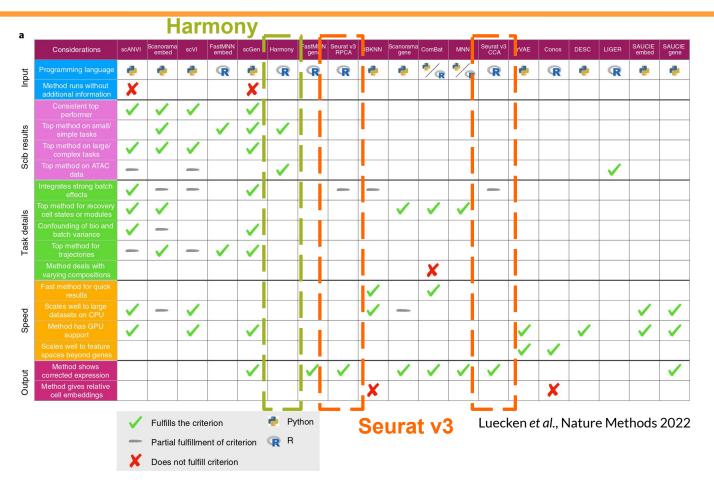
Diagonal: different samples different modalities



Many methods



Many methods



A few benchmarks, that do not agree with each other

Büttner et al., Nat. Methods. 2019 Chen et al., Nat. Biotechnol 2020 Tran et al., Genome Biol. 2020

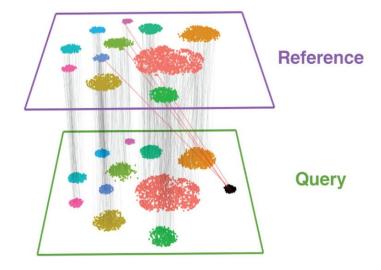
Integration with Seurat



Algorithm



- MNN: Mutual Nearest Neighbors
- In reference and query, identify 2 cells that are close (neighbors) in terms of euclidean distance: **anchors**
- Identify many anchors





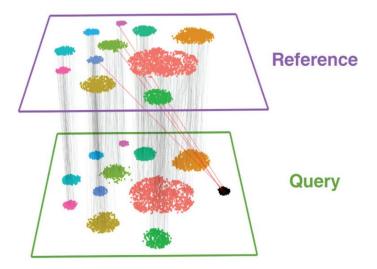
Integration with Seurat



Algorithm



- Deduce correction from anchors
- Apply correction vector to all query cells.





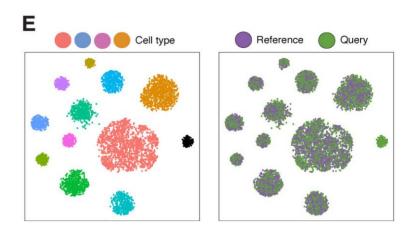
Integration with Seurat



Algorithm



- Deduce correction from anchors
- Apply correction vector to all query cells.





Conclusion

A good integration method

Technical

Biological



- Corrects for technical variability:
 - samples
 - donors
 - experimenter
 - technologies

- Preserves biological signal
 - cell types across different samples, tissues
 - cell trajectories
 - differences (cell subtypes, cell states) between condition and control
 - population (cell subtypes, cell states) unique to a condition...

Acknowledgements

Parts of this course are inspired by

The Swiss Institute of Bioinformatics course Single Cell Transcriptomics

Slides inspired from Rémi Montagne (Thanks!)