



alliance nationale pour les sciences de la vie et de la santé

Processing 1 Normalization, scaling and regression

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Organisation of the scRNA-seq course

- From raw count matrix to normalised matrix
 - Filtering low quality droplets
 - Filtering dead cells
 - Filtering doublets
- Data normalization
 - Why do we need to normalize the data ?
 - What are the methods available ?
 - Regression of biological biaises

Question!

What do you think of each gene expression in this matrix ?

| Gène | Cellule 1 | Cellule 2 | Cellule 3 | Cellule 4 |
|-------|-----------|-----------|-----------|-----------|
| Rouge | 100 | 200 | 300 | 400 |
| Bleu | 50 | 100 | 150 | 200 |
| Vert | 10 | 10 | 10 | 10 |
| Jaune | 100 | 100 | 100 | 100 |

Why do we need to normalize our data?

We need to remove **technical biases** in order to...

Condition A : 12 reads



Images from https://www.biostars.org/p/349881/

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Condition A : 12 reads

The 2 libraries have the **same RNA composition**.

But the condition B has 3 times more reads than the condition A.

We need to correct for differences in **library size**.

Images from https://www.biostars.org/p/349881/

Why do we need to normalize our data?

We need to remove **technical biases** in order to...

Normalization allows us to compare cells



What do you know about normalization ?

How many approaches do you think there are ?

Plenty of normalization approaches for bulk RNA-seq

- TPM
- CPM
- RPKM
- FPKM
- Global scaling (eg: Upper Quartile)
- Size factors calculation (eg: estimation of library sampling depth) :
 - DESeq2
 - \circ edgeR
- ...

Plenty of normalization approaches for bulk RNA-seq

- TPMCPM
 - ×
- RPKM

•

FPKM



These methods do not apply to single-cell data (or partially)

• Global scaling (eg: Upper Quartile)



- Size factors calculation (eg: estimation of library sampling depth) :
 - DESeq2
 - edgeR





This is mostly due to the sparsity of the single-cell data



A sparse matrix is a matrix filled with a LOT of zeros

First, let's take this simple raw count matrix :

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Our first impressions are like :

Gene rouge : expression is proportional to the sequencing depth Gene bleu : same as "gene rouge" but with smaller values Gene vert : expression is low and steady Gene jaune : expression is steady between the cells

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 Sequencing depth :
 260
 410
 560
 710

CPM normalized value for gene rouge in cellule 1 : 100/260*10⁴= 6250

Let's apply this to all genes :

| Gène | Cellule 1 | Cellule 2 | Cellule 3 | Cellule 4 |
|-------|-----------|-----------|-----------|-----------|
| Rouge | 6250 | 6451 | 6522 | 6557 |
| Bleu | 3125 | 3226 | 3261 | 3279 |
| Vert | 625 | 323 | 217 | 164 |
| Jaune | 6250 | 3226 | 2174 | 1639 |

After CPM normalization, we have :

Gene rouge : the expression is actually steady between the cells Gene bleu : the expression is actually steady between the cells (but lower than gene rouge) Gene vert : expression decreases Gene jaune : expression decreases

We applied this formula (we just omitted the log and the pseudo count "+1") :

$$ext{Normalized value}_{g,c} = \log_2\left(rac{ ext{UMI}_{g,c}}{ ext{TotalUMI}_c} imes ext{ScaleFactor} + 1
ight)$$

Why do we add the log transformation ?

- To reduce the impact of extreme values
- To stabilize the variance
- To have a better biological interpretation
- Improved Model Fit



What about scaling ???

Why do we scale ?

• To be able to improve comparability between genes



Examples of biological biases that you may want to correct :

- Amplification
- RNA capture efficiency
- Gene length
- GC content

How do we scale?

Let's scale gene rouge :

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Mean expression of gene rouge = 25780 / 4 = 6445 Standard deviation $\sigma \approx 137.3$

How do we scale?

Let's scale gene rouge :

Mean expression of gene rouge = 25780 / 4 = 6445 Standard deviation $\sigma \approx 137.3$

Use the formula:

$$\mathrm{Scaled_value}_i = rac{\mathrm{Value}_i - \mu}{\sigma}$$

Calculations for each cell:

For 6250 :
$$\frac{6250 - 6445}{137.3} = \frac{-195}{137.3} \approx -1.42$$

For 6451 : $\frac{6451 - 6445}{137.3} = \frac{6}{137.3} \approx 0.04$
For 6522 : $\frac{6522 - 6445}{137.3} = \frac{77}{137.3} \approx 0.56$
For 6557 : $\frac{6557 - 6445}{137.3} = \frac{112}{137.3} \approx 0.82$

This is the z-score !

- Scaling ensures that the values have a mean of 0 and a standard deviation of 1
- This transformation makes genes with different ranges comparable across cells or datasets

Visualisation of the impact of each transformation



Be careful ! Scaling may not always be necessary...

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For example:

- In differential expression analysis, raw or log-normalized counts are used since absolute expression levels matter.
- For some downstream analyses like gene set enrichment analysis (GSEA), raw expression values are often more meaningful.

HO NO !

- Some close relative offered you your favorite gear : a nice T-shirt with a cool design
- Unfortunately your 5-years old inadvertently tainted it with its paint flask...
- Hopefully, you know a skilled team of cleaners right around the corner (who actually are bioinformaticians in disguise ! *wink wink*)
- They explain you they learned how to clean stains by **accumulating knowledge**, having cleaning tons of them in several contexts.

Cleaners to the rescue

• By compiling this amount of knowledge, they could produce a specific cleaning protocol to the kind of staining affecting your Tee, subtracted the former to the latter, revealing the object of desire

Let's get back to reality

• This is what is performed in the cell cycle example, thanks to acquired data on the cell-cycle -specific gene expression, and regression

The devil may be in the bottle

- But what would happen if by bad luck, the cleaner team trained themselves on stains that unfortunately *look a bit to much like* your pattern ?
- The same way you **increased signal by removing some noise** on the former example, one may perform the **complete opposite** if one doesn't pay attention
- Probably because in this specific context, what you thought was noise, wasn't

Acknowledgements

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