# Intra-specific and interspecific protein content analysis

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École "Assemblage & Annotation" AVIESAN 2022 - Roscoff

#### Intra-specific and interspecific protein content analysis

Goals :

- Reduce redondancy (alternative CDS) => compute representative sequences
- Build set of repeated sequences or highly conserved families(genes/proteins)
- Compare gene/protein content between
  - o assemblies versions
  - individuals of a single species
  - o different species

=> Gene loss/gain (toward pangenomics)

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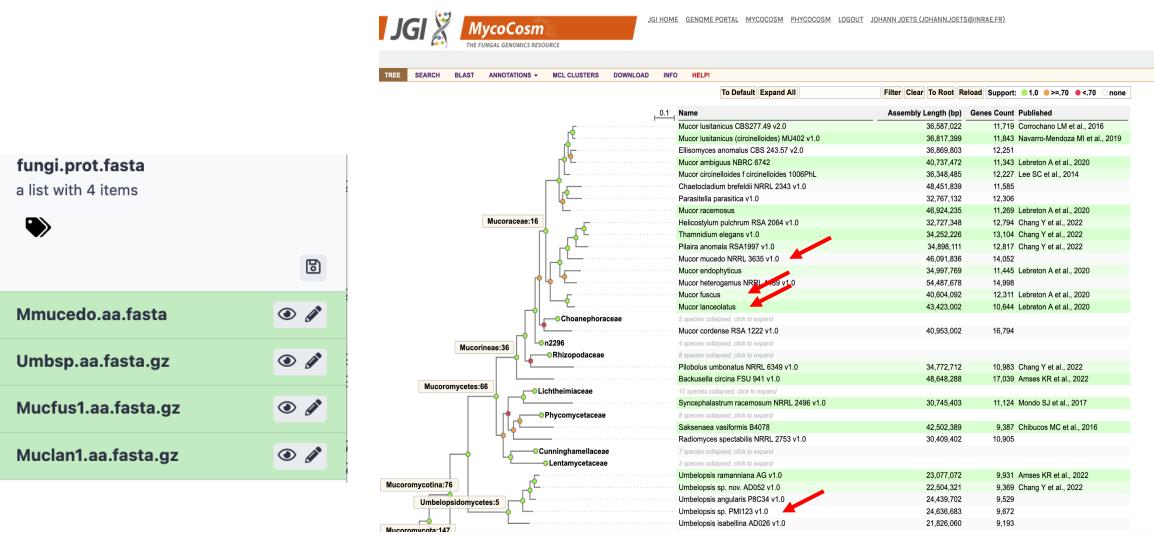
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Methods / tools :

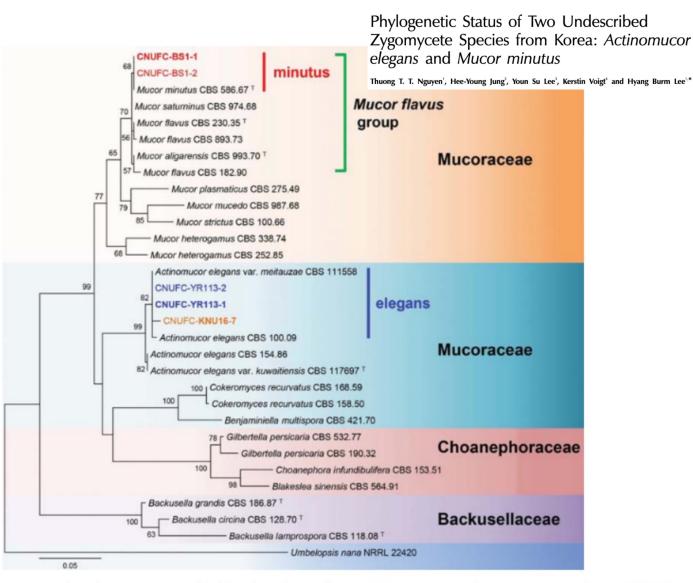
- Sequence similarity-based clustering : **CD-HIT** => intra-specific comparaison (*low seq divergence*)
- Phylogeny-based orthology classification : **OrthoFinder** => inter-specific comparaison (*higher seq divergence*)

## **OrthoFinder. Our dataset:**



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**Fig. 2.** Phylogenetic tree based on maximum likelihood analysis of 28S rDNA sequences for *Actinomucor elegans* CNUFC-YR113-1, *A. elegans* CNUFC-YR113-2, *A. elegans* CNUFC-KNU16-7, *Mucor minutus* CNUFC-BS1-1, and *M. minutus* CNUFC-BS1-2. *Umbelopsis nana* was used as an outgroup. Bootstrap support values of  $\geq$  50% are indicated at the nodes. The bar indicates the number of substitutions per position.



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# **OrthoFinder: Galaxy**

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All workflows	<ul> <li>24: OrthoFinder on data 5, data 6, and others: overall comparative genomics statistics</li> <li>25: OrthoFinder on data 5, data 6, and others: per species comparative genomics statistics</li> <li>26: OrthoFinder on data 5, data 6, and others: species tree</li> <li>27: OrthoFinder on data 5, data 6, and others: species tree with node labels</li> <li>28: OrthoFinder on data 5, data 6, and others: species tree with duplication events</li> <li>29: OrthoFinder on data 5, data 6, and others: duplication events</li> <li>30: OrthoFinder on data 5, data 6, and others: duplications per orthogroup</li> <li>31: OrthoFinder on data 5, data 6, and others: duplications per species tree node</li> </ul> You can check the status of queued jobs and view the resulting data by refreshing the History panel. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.								

# CD-HIT a sequence clustering program

Was originaly developed to remove redundancy and select representative sequences at verry high speed

CD-HIT, use incremental clustering algorithm method:

- 1/ Sequences are sorted in order of decreasing length.
- 2/ The longest one becomes the representative of the first cluster.
- 3/ Each remaining sequence is compared to the representatives of existing clusters.
  - If the similarity with any representative is above a given threshold, it is grouped into that cluster.
  - Otherwise, a new cluster is defined with that sequence as the representative.

Alternatives transcripts

Painfull for gene content comparison between genomes.

Try to select a representative sequence per gene.

#### 📕 Galaxy France

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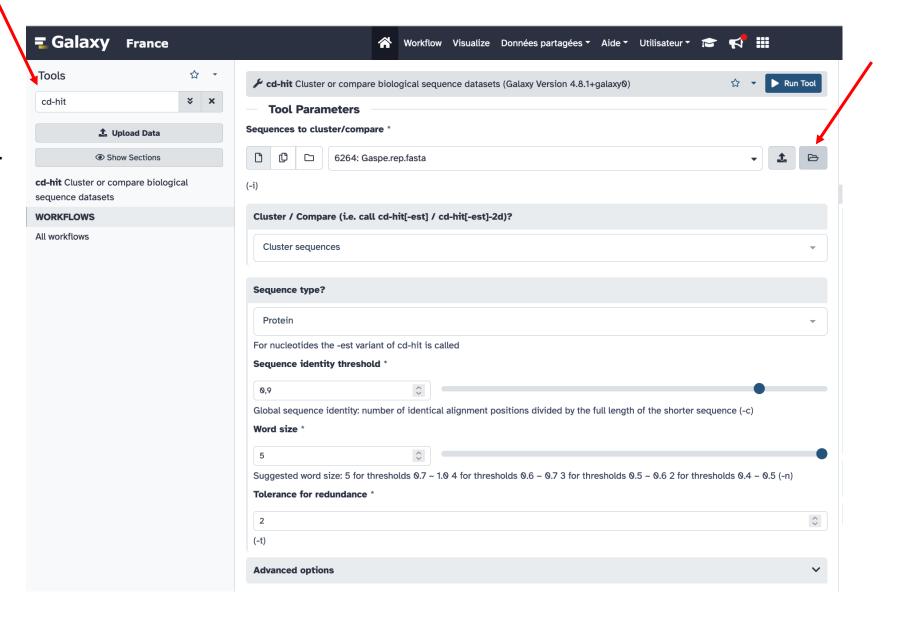
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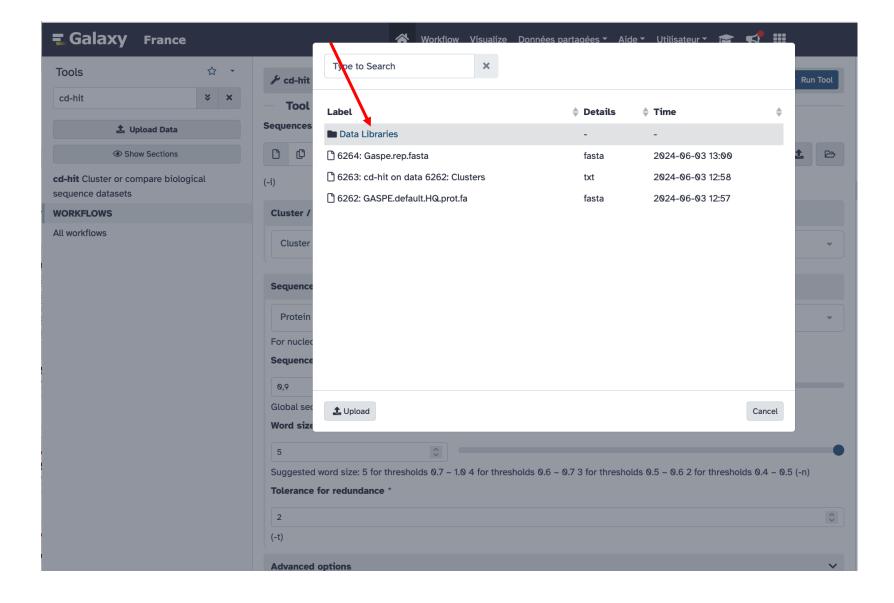
Alternatives transcripts

Painfull for gene content comparison between genomes.



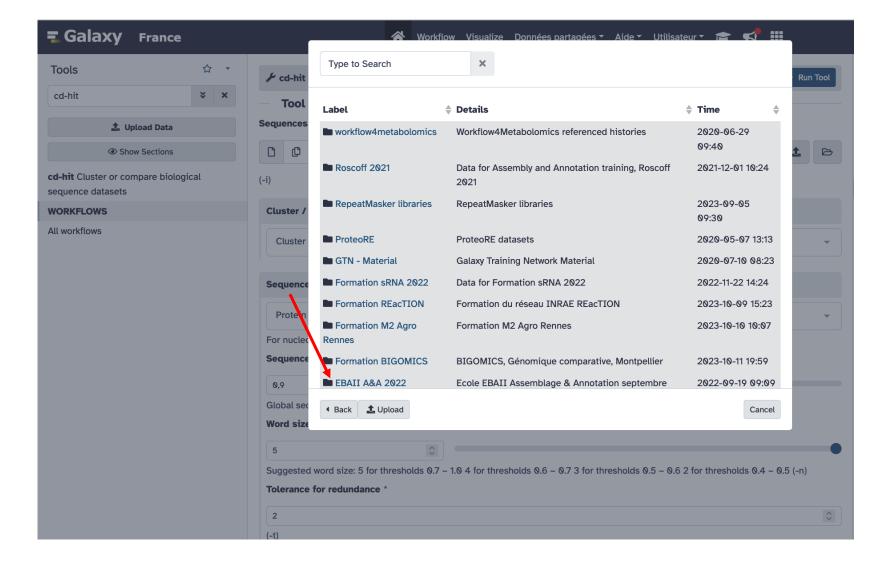
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Alternatives transcripts

Painfull for gene content comparison between genomes

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Alternatives transcripts

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Try to select a representative sequence per gene.

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>Cluster 29			
0 110aa, >Zm00062aa026574_T00 at 99.09%			
1 2552aa, >Zm00062aa033122_T00 *			Program: CD-HIT, V4.8.1 (+OpenMP), Aug 07
>Cluster 30			2022, 06:48:22
0 2538aa, >Zm00062aa015588_T00 *			2022, 00.40.22
>Cluster 31			🖬 🔗 🚯 C' 📖 🊠 ?
0 2478aa, >Zm00062aa013952_T00 *			>Cluster 0
>Cluster 32 0 2449aa. >Zm00062aa035739 T00 *			
0 2449aa, >Zm00062aa035739_T00 * >Cluster 33			0 5422aa, >Zm00062aa029250_T00 at 10
0 2447aa, >Zm00062aa035984_T00 *			1 5425aa, >Zm00062aa029250_T00 *
1 1604aa, >Zm00062aa035984_T00 at 99.31%			>Cluster 1
>Cluster 34			0 5066aa, >Zm00062aa031031_T00 *
0 2441aa, >Zm00062aa042116_T00 *			
>Cluster 35			6265: F331.default.HQ.prot.fa 🛛 💿 🧪 🥤
0 2439aa, >Zm00062aa028764_T00 *			
>Cluster 36 0     2410aa, >Zm00062aa008109_T00 at 100.00%			6264: Gaspe.rep.fasta 🛛 💿 🧪 📋
1 2410aa, >Zm00062aa008109_T00 at 100.00%			
2 2410aa, >Zm00062aa008109_T00 at 100.00%			6263: cd-hit on data 6262: Clust 🛛 🖉 📋
3 2410aa, >Zm00062aa008109_T00 at 100.00%			ers
4 2432aa, >Zm00062aa008109_T00 *			
>Cluster 37			

(\*) Sequence selected as representative

Alternatives transcripts

Painfull for gene content comparison between genomes.

Try to select a representative sequence per gene.

This dataset is large and only the first megabyte is shown below.	Rechercher des données	*
Show all   Save		
Zm00062aa000001_T001	data	
HANMPWIMHAGTEQRHAACGASLLWSSLQPSTVVMAAAAATFGFLHPPIRKPAVPPLYILRLPTKPHSK		
HPRSPPLLFLLLGRRRGGPIAAFPNTTSSSTNAPASPTYDVREAEAAVADLLREGGASADDAASIAARA AYAAMLADGVRELDELGLWASWSSGARARLGLSGVVEMEMGRLGFRRKVYLMGRSKPDHGVVPLLESLG	<b>86.3 MB</b>	6 🗑 6.
RLSSAKLIAPYVAAAGLTVLIDRVKFLKEMLFSSSDYAILIGRNAKRMMTYLSIPADDALQSTLSFFEK	<b>2</b> 49	
EARYGGVSMLGHGDVSFPYLIESFPMLLLCSEDNHLKPLVDFLEHIGIPKPKIASVLLLFPPIILSDVE	<b>M</b>	4
DIKPRIREWEKAGIEQDYVSRMLLKYPWILSTSVIENYSQMLLFFNQKRISSTVLAIAVKSWPHILGSS	6267: F331.rep.fasta	l l l l l l l l l l l l l l l l l l l
<pre>KRMNSVLELFHVLGISKKMVVPVITSSPQLLLRKPDQFMQNVLFFREMGVDKKTTGKILCRSPEIFASN</pre>		L C
DNTLKKKIDFLTNFGVSKHHLPRIIRKYPELLLLDINCTLLPRMNYLLEMGLSKKDLCSMIFRFSPLLG	6266: cd-hit on data 626	5: Clust 🤇
SIELVMKPKLEFLLRTMKKPLKAVVEYPRYFSYSLEGKIKPRFWVLQSRNIDCTLTEMLAKNDELFAEE	ers	
_GLGGLLEKPLQSSIGS Zm00062aa000002_T001		
-YLCSSVCCSIPSRSLSFSFYFFVTILNDNYRKHGERPPQPPLEIVKYPALLERAWGWDSILPYCPVNS	6265: F331.default.HQ.pr	ot.fa 🤇
PLYKTYLQEYYERNSREVLAHQVEANAASGPNLNGADENDLATLFNLCISREVQLLNLLKRRDQKHNVD		
ISLNEKLTNCARQITFVEYAGFPFPSVALKCIMAESDLLCMLLMCGTGMISQVNICSRIRKVAFRFMTY	6264: Gaspe.rep.fasta	
JPGCFAAAATMMATTKEAKLMSGLLRNRCRENNGPFSRSVFIRKWTIAAMFRICEECSPVEESTGGYTA		
KLILDGSDDKNPCDKELVNKDNLLQRNKINQKYQGLFKGKTTKCRHPVQKQEPGDGATPASPSPANPIS VVOKLW	6263: cd-hit on data 626	2: Clust 🤇
Zm00062aa000002_T002	ers	
AMAEKVKEKMLMLRSSDNQEFEVKESVAMQSMTLKKMVEDGCADKGIPLPNVTSHILVKVIEYCNKHAE		
TGPGDAAGTTNRSAEDELNIFDADFVNVEHSTLLDLILAANYLDIKGLLNLARQTITDLINGKMPEEVC	6262: GASPE.default.HQ.	prot.fa 🧕 🥝
TNIKNDLTIPSTSALATTMPSSERKQMEARFIAYMQETAEERMTGFAVRGEKYRKHGERPPQPPLEIVK		
PALLERAWGWDSILPYCPVNSWPLYKTYLQEYYERNSREVLAHQVEANAASGPNLNGADENDLATLFNL		
ISREVQLLNLLKRRDQKHNVDEISLNEKLTNCARQITFVEYAGFPFPSVALKCIMAESDLLCMLLMCGT MISQVNICSRIRKVAFRFMTYKGPGCFAAAATMMATTKEAKLMSGLLRNRCRENNGPFSRSVFIRKWTI		
MISQUNICSKIKKVAFKFMIIKQPGCFAAAAIMMATIKEAKLMSGLLKNKCKENNGPFSKSVFIKKWII AMFRICEECSPVEESTGGYTATKLILDGSDDKNPCDKELVNKDNLLQRNKINQKYQGLFKGKTTKCRHP		
QKQEPGDGATPASPSPANPISTVVQKLW		
Zm00062aa000002_T004		
AMAEKVKEKMLMLRSSDNQEFEVKESVAMQSMTLKKMVEDGCADKGIPLPNVTSHILVKVIEYCNKHAE		
TGPGDAAGTTNRSAEDELNIFDADFVNVEHSTLLDLILAANYLDIKGLLNLARQTITDLINGKMPEEVC		
TNIKNDLTIPSTSALATTMPSSERKQMEARFIAYMQETAEERMTGFAVRGEKYRKHGERPPQPPLEIVK		
PESMVNVRHNHHWR Zm00062aa000003_T001		
PSALRRLFATILVYCEPSDVAVLWOKHLDAMSEDYORRSOSKTHVEOMVLIDIRNMLOSMGKDIKTFPL		
PIIDAYDDAIGTAREVYEEESIEPAAGDVALKDSLNEEQRAAYDKILSAVDTDQGGLFFVDGPGGTEKT		
LYRVPLTTLRSQGKIAVATATSGVAASIMPGGRTAHSRFKIPLTIDDGAVLPVVRKGSRAQVVASSLWM		
YLWESMSHLKLVSNMRTKNDPWFAEYLLRVGGGTEVTNSDGDIRLPDEVCVPYSGSDSDLDNLIDFVFP		
LNENMSDSTYITSRAILSTRNDWVDMINAKMIDRFQGEHTVYHSFDSAMDDPHNYYPPEFLNTLTPNGL		

🗥 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 定 📢 🏢

Using 12%

Alternatives transcripts

Painfull for gene content comparison between genomes.

Try to select a representative sequence per gene.

This dataset is large and only the first megabyte is shown below.	Rechercher des données	*
Show all   Save		
Zm00062aa000001_T001	data	
HANMPWIMHAGTEQRHAACGASLLWSSLQPSTVVMAAAAATFGFLHPPIRKPAVPPLYILRLPTKPHSK		
HPRSPPLLFLLLGRRRGGPIAAFPNTTSSSTNAPASPTYDVREAEAAVADLLREGGASADDAASIAARA AYAAMLADGVRELDELGLWASWSSGARARLGLSGVVEMEMGRLGFRRKVYLMGRSKPDHGVVPLLESLG	<b>86.3 MB</b>	6 🗑 6.
RLSSAKLIAPYVAAAGLTVLIDRVKFLKEMLFSSSDYAILIGRNAKRMMTYLSIPADDALQSTLSFFEK	<b>2</b> 45	
EARYGGVSMLGHGDVSFPYLIESFPMLLLCSEDNHLKPLVDFLEHIGIPKPKIASVLLLFPPIILSDVE	<b>M</b>	4
DIKPRIREWEKAGIEQDYVSRMLLKYPWILSTSVIENYSQMLLFFNQKRISSTVLAIAVKSWPHILGSS	6267: F331.rep.fasta	l l l l l l l l l l l l l l l l l l l
<pre>KRMNSVLELFHVLGISKKMVVPVITSSPQLLLRKPDQFMQNVLFFREMGVDKKTTGKILCRSPEIFASN</pre>		L C
DNTLKKKIDFLTNFGVSKHHLPRIIRKYPELLLLDINCTLLPRMNYLLEMGLSKKDLCSMIFRFSPLLG	6266: cd-hit on data 626	5: Clust 🤇
SIELVMKPKLEFLLRTMKKPLKAVVEYPRYFSYSLEGKIKPRFWVLQSRNIDCTLTEMLAKNDELFAEE	ers	
_GLGGLLEKPLQSSIGS Zm00062aa000002_T001		
-YLCSSVCCSIPSRSLSFSFYFFVTILNDNYRKHGERPPQPPLEIVKYPALLERAWGWDSILPYCPVNS	6265: F331.default.HQ.pr	ot.fa 🤇
PLYKTYLQEYYERNSREVLAHQVEANAASGPNLNGADENDLATLFNLCISREVQLLNLLKRRDQKHNVD		
ISLNEKLTNCARQITFVEYAGFPFPSVALKCIMAESDLLCMLLMCGTGMISQVNICSRIRKVAFRFMTY	6264: Gaspe.rep.fasta	
JPGCFAAAATMMATTKEAKLMSGLLRNRCRENNGPFSRSVFIRKWTIAAMFRICEECSPVEESTGGYTA		
KLILDGSDDKNPCDKELVNKDNLLQRNKINQKYQGLFKGKTTKCRHPVQKQEPGDGATPASPSPANPIS VVOKLW	6263: cd-hit on data 626	2: Clust 🤇
Zm00062aa000002_T002	ers	
AMAEKVKEKMLMLRSSDNQEFEVKESVAMQSMTLKKMVEDGCADKGIPLPNVTSHILVKVIEYCNKHAE		
TGPGDAAGTTNRSAEDELNIFDADFVNVEHSTLLDLILAANYLDIKGLLNLARQTITDLINGKMPEEVC	6262: GASPE.default.HQ.	prot.fa 🧕 🥝
TNIKNDLTIPSTSALATTMPSSERKQMEARFIAYMQETAEERMTGFAVRGEKYRKHGERPPQPPLEIVK		
PALLERAWGWDSILPYCPVNSWPLYKTYLQEYYERNSREVLAHQVEANAASGPNLNGADENDLATLFNL		
ISREVQLLNLLKRRDQKHNVDEISLNEKLTNCARQITFVEYAGFPFPSVALKCIMAESDLLCMLLMCGT MISQVNICSRIRKVAFRFMTYKGPGCFAAAATMMATTKEAKLMSGLLRNRCRENNGPFSRSVFIRKWTI		
MISQUNICSKIKKVAFKFMIIKQPGCFAAAAIMMATIKEAKLMSGLLKNKCKENNGPFSKSVFIKKWII AMFRICEECSPVEESTGGYTATKLILDGSDDKNPCDKELVNKDNLLQRNKINQKYQGLFKGKTTKCRHP		
QKQEPGDGATPASPSPANPISTVVQKLW		
Zm00062aa000002_T004		
AMAEKVKEKMLMLRSSDNQEFEVKESVAMQSMTLKKMVEDGCADKGIPLPNVTSHILVKVIEYCNKHAE		
TGPGDAAGTTNRSAEDELNIFDADFVNVEHSTLLDLILAANYLDIKGLLNLARQTITDLINGKMPEEVC		
TNIKNDLTIPSTSALATTMPSSERKQMEARFIAYMQETAEERMTGFAVRGEKYRKHGERPPQPPLEIVK		
PESMVNVRHNHHWR Zm00062aa000003_T001		
PSALRRLFATILVYCEPSDVAVLWOKHLDAMSEDYORRSOSKTHVEOMVLIDIRNMLOSMGKDIKTFPL		
PIIDAYDDAIGTAREVYEEESIEPAAGDVALKDSLNEEQRAAYDKILSAVDTDQGGLFFVDGPGGTEKT		
LYRVPLTTLRSQGKIAVATATSGVAASIMPGGRTAHSRFKIPLTIDDGAVLPVVRKGSRAQVVASSLWM		
YLWESMSHLKLVSNMRTKNDPWFAEYLLRVGGGTEVTNSDGDIRLPDEVCVPYSGSDSDLDNLIDFVFP		
LNENMSDSTYITSRAILSTRNDWVDMINAKMIDRFQGEHTVYHSFDSAMDDPHNYYPPEFLNTLTPNGL		

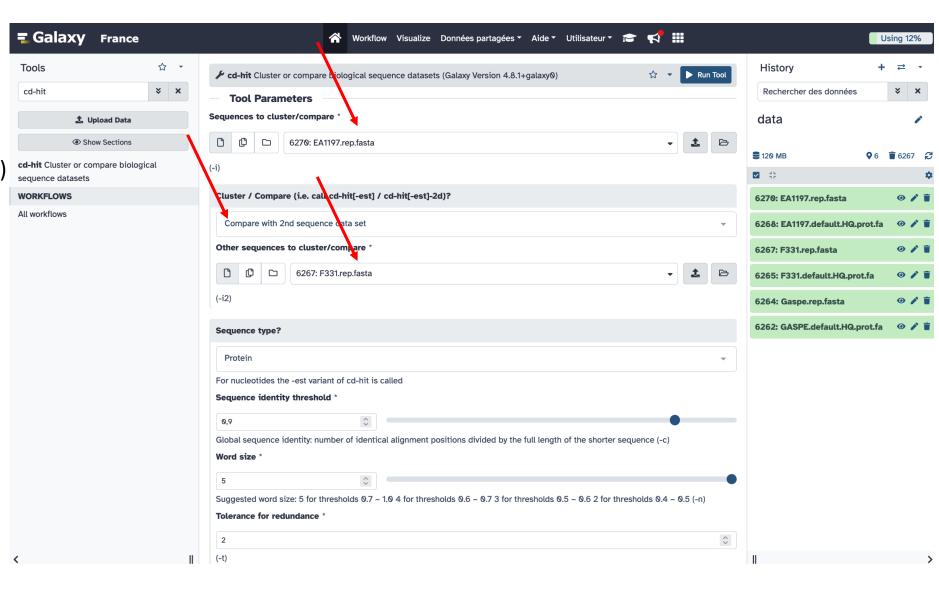
🗥 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 定 📢 🏢

Using 12%

Compare 2 gene/protein dataset

From 2 annotation versions From 2 individuals (same species)

Input : outputs from use-case 1



Compare 2 gene/protein dataset

From 2 annotation versions

Zm00063 : EA1197 Zm00062 : F331

```
This dataset is large and only the first megabyte is shown below.
       Show all | Save
>Cluster 0
        5425aa, >Zm00063aa018611_T00... *
0
1
        5425aa, >Zm00062aa029250_T00... at 99.83%
        5066aa, >Zm00063aa032449_T00... *
1
        5066aa, >Zm00062aa031031_T00... at 99.98%
>Cluster 2
0
        4863aa, >Zm00063aa019422_T00... *
1
        4863aa, >Zm00062aa006742_T00... at 99.40%
>Cluster 3
0
        4210aa, >Zm00063aa003366_T00... *
1
        4210aa, >Zm00062aa023384_T00... at 99.95%
>Cluster 4
0
        3882aa, >Zm00063aa034457_T00... *
        3877aa, >Zm00062aa033117_T00... at 99.95%
1
>Cluster 5
0
        3691aa, >Zm00063aa030400_T00... *
        3691aa, >Zm00062aa005734_T00... at 99.78%
1
>Cluster 6
        3650aa, >Zm00063aa035322_T00... *
0
        3631aa, >Zm00062aa013354_T00... at 92.29%
1
        3642aa, >Zm00062aa041101_T00... at 99.84%
2
>Cluster 7
        3631aa, >Zm00063aa007786_T00... *
0
>Cluster 8
0
        3581aa, >Zm00063aa030440_T00... *
1
        3581aa, >Zm00062aa005775_T00... at 99.83%
>Cluster 9
0
        3287aa, >Zm00063aa002573_T00... *
>Cluster 10
        3212aa, >Zm00063aa015366_T00... *
0
>Cluster 11
        3007aa, >Zm00063aa008396_T00... *
0
>Cluster 12
0
        2872aa, >Zm00063aa020205_T00... *
```

History	+	₽	•	
Rechercher des données		*	×	
data			-	
130 MB	<b>9</b> 8	626	67 🖌	3
2 11				¢
6272: cd-hit on data 6267 ta 6270: Representative se es			/1	
6271: cd-hit on data 6267 a ta 6270: Clusters	and da	0	1	ī
6270: EA1197.rep.fasta		0	1	ī
6268: EA1197.default.HQ.p	rot.fa	0	1	ī
6267: F331.rep.fasta		0	1	ī
6265: F331.default.HQ.prot	.fa	0	1	ī
6264: Gaspe.rep.fasta		0	1	ī
6262: GASPE.default.HQ.pr	rot.fa	0	1	ī

Using 12%

Compare 2 gene/protein dataset

From 2 annotation versions From 2 individuals (same species)

Zm00063 : EA1197 Zm00062 : F331

```
This dataset is large and only the first megabyte is shown below.
       Show all | Save
>Cluster 0
        5425aa, >Zm00063aa018611_T00... *
0
1
        5425aa, >Zm00062aa029250_T00... at 99.83%
        5066aa, >Zm00063aa032449_T00... *
        5066aa, >Zm00062aa031031_T00... at 99.98%
1
>Cluster 2
0
        4863aa, >Zm00063aa019422_T00... *
        4863aa, >Zm00062aa006742_T00... at 99.40%
1
>Cluster 3
        4210aa, >Zm00063aa003366_T00... *
0
        4210aa, >Zm00062aa023384_T00... at 99.95%
1
>Cluster 4
0
        3882aa, >Zm00063aa034457_T00... *
        3877aa, >Zm00062aa033117_T00... at 99.95%
1
>Cluster 5
0
        3691aa, >Zm00063aa030400_T00... *
        3691aa, >Zm00062aa005734_T00... at 99.78%
1
>Cluster 6
        3650aa, >Zm00063aa035322_T00... *
                                                                  Orthologous gene to 13354 is
0
        3631aa, >Zm00062aa013354_T00... at 92.29%
1
                                                                  missing in EA1197?
        3642aa, >Zm00062aa041101_T00... at 99.84%
2
>Cluster 7
0
        3631aa, >Zm00063aa007786_T00... *
>Cluster 8
0
        3581aa, >Zm00063aa030440_T00... *
1
        3581aa, >Zm00062aa005775_T00... at 99.83%
>Cluster 9
0
        3287aa, >Zm00063aa002573_T00... *
>Cluster 10
        3212aa, >Zm00063aa015366_T00... *
0
>Cluster 11
        3007aa, >Zm00063aa008396_T00... *
0
>Cluster 12
0
        2872aa, >Zm00063aa020205_T00... *
```

Workflow Visualize Données partagées - Aide - Utilisateur -

History +	₽	•
Rechercher des données	*	×
data		1
€ 130 MB	626	7 2
M at		•
6272: cd-hit on data 6267 and da ta 6270: Representative sequenc es	0	/ 1
6271: cd-hit on data 6267 and da ta 6270: Clusters	0	/ 1
6270: EA1197.rep.fasta	0	/ 1
6268: EA1197.default.HQ.prot.fa	0	/ 1
6267: F331.rep.fasta	0	/ 1
6265: F331.default.HQ.prot.fa	0	/ 1
6264: Gaspe.rep.fasta	0	/ 1
6262: GASPE.default.HQ.prot.fa	0	/ 1

Using 12%

Orthologous genes missing in F331?

Compare n gene/protein dataset

From n annotation versions From n individuals (same species)

Input : outputs from UC1

Step 1 files concatenation

<b>=</b> Galaxy France	ilisateur - 😭 👫 🗰	
Tools 🔄 🔹	✓ Concatenate datasets tail-to-head (cat) (Galaxy Version 9.3+galaxy1) ☆ & ▼ ▶ Tool Parameters	Run Tool
1 Upload Data	Datasets to concatenate *	
<ul> <li>Show Sections</li> <li>Concatenate datasets tail-to-head (cat)</li> <li>Concatenate datasets tail-to-head</li> <li>Concatenate FASTA alignment by species</li> <li>bcftools concat Concatenate or combine</li> </ul>	6272: cd-hit on data 6267 and data 6270: Representative sequences         6271: cd-hit on data 6267 and data 6270: Clusters         6270: EA1197.rep.fasta         6268: EA1197.default.HQ.prot.fa         6265: F331.default.HQ.prot.fa         6265: F331.default.HQ.prot.fa	
VCF/BCF files Concatenate two BED files hamronize summarize: Concatenate and	1: Dataset	Î
summarize AMR detection reports Rename.seqs Rename sequences by concatenating the group name FASTA Merge Files and Filter Unique Sequences Concatenate FASTA database files together	Select * 6272: cd-hit on data 6267 and data 6270: Representative sequences 6271: cd-hit on data 6267 and data 6270: Clusters 6270: EA1197.rep.fasta 6268: EA1197.default.HQ.prot.fa 6265: F331.rep.fasta 6265: F331.default.HQ.prot.fa 6265: F331.default.HQ.prot.fa	
WORKFLOWS		-
All workflows	2: Dataset Select * C279: EATT97.rep.tasta 6268: EAT197.default.HQ.prot.fa 6267: F331.default.HQ.prot.fa 6265: F331.default.HQ.prot.fa 6262: GASPE.default.HQ.prot.fa 6262: GASPE.default.HQ.prot.fa	
	+ Insert Dataset	

Compare n gene/protein dataset

From n annotation versions From n individuals (same species)

Input : outputs from UC1

Step 1 files concatenation

Step 2 cluster seq with CD-Hit

<b>= Galaxy</b> France	🎢 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 🞓 📢 🏢
Tools ☆ •	🗲 cd-hit Cluster or compare biological sequence datasets (Galaxy Version 4.8.1+galaxy0) 🏠 🔹 🕨 Run Tool
cd-hit × ×	Tool Parameters Sequences to cluster/compare *
Show Sections	Image: Selected: /Comparative genomics/F331.Gaspe.EA1197.rep.fasta
<b>cd-hit</b> Cluster or compare biological sequence datasets	(-i)
WORKFLOWS	Cluster / Compare (i.e. call cd-hit[-est] / cd-hit[-est]-2d)?
All workflows	Cluster sequences -
	Sequence type?
	Protein -
	For nucleotides the -est variant of cd-hit is called
	Sequence identity threshold *
0.97	0,97 Clobal sequence identity: number of identical alignment positions divided by the full length of the shorter sequence (-c)

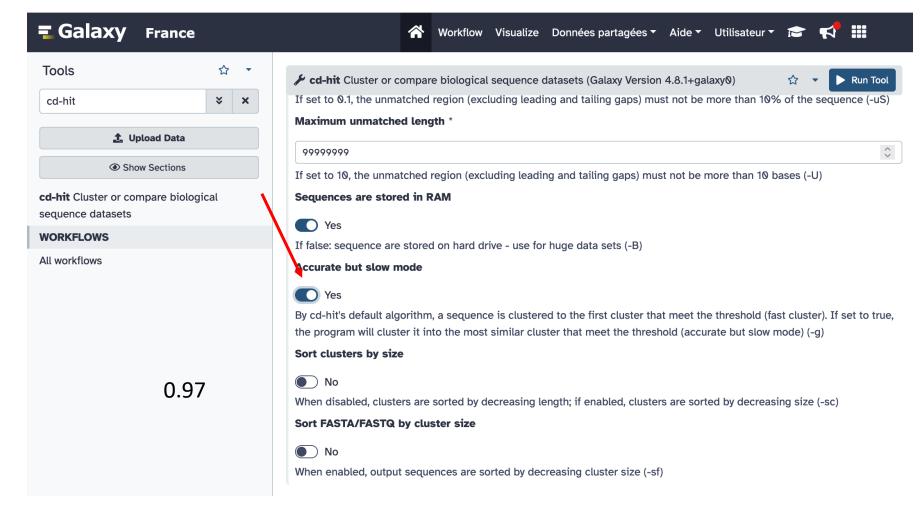
#### Compare n gene/protein dataset

From n annotation versions From n individuals (same species)

Input : outputs from UC1

Step 1 files concatenation

Step 2 cluster seq with CD-Hit



CD-HIT Use case 3 : e	<b>=</b> Galaxy France	isualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 🞓 📢 🏭	
Compare n gene/prot	Tools ☆ ▼ cd-hit × ×	0 130aa, >Zm00063aa043711_T00 * >Cluster 60001 0 130aa, >Zm00063aa044627_T00 *	F
From n annotation ve From n individuals (sa	1 Upload Data	1 130aa, >Zm00062aa040294_T00 at 100.00% >Cluster 60002 0 130aa, >Zm00063aa044846_T00 * >Cluster 60003 0 130aa, >Zm00063aa044897_T00 *	C
Input : outputs from L	<ul> <li>Show Sections</li> <li>cd-hit Cluster or compare biological sequence datasets</li> </ul>	1 130aa, >Zm00062aa040567_T00 at 100.00% 2 130aa, >Zm00064aa040891_T00 at 100.00% >Cluster 60004 0 130aa, >Zm00062aa000798_T00 *	€ 2
Step 1 files concatena	WORKFLOWS All workflows	>Cluster 60005 0 130aa, >Zm00062aa001058_T00 * 1 130aa, >Zm00064aa001104_T00 at 100.00% >Cluster 60006 0 130aa, >Zm00062aa002134_T00 *	3: ve
Step 2 cluster seq witl		<pre>&gt;Cluster 60007 0 130aa, &gt;Zm00062aa004172_T00 * 1 130aa, &gt;Zm00064aa004266_T00 at 100.00% &gt;Cluster 60008</pre>	2: 1:
Zm00064 : Gaspe Zm00063 : EA1197		0 130aa, >Zm00062aa004378_T00 * 1 130aa, >Zm00064aa004472_T00 at 100.00% >Cluster 60009 0 130aa, >Zm00062aa005951_T00 * 1 130aa, >Zm00064aa006122_T00 at 99.23% >Cluster 60010 0 130aa = Zm00062aa006314_T00	
Zm00062 : F331		0 130aa, >Zm00062aa006214_T00 * 1 107aa, >Zm00064aa006390_T00 at 100.00% >Cluster 60011 0 130aa, >Zm00062aa006445_T00 * >Cluster 60012 0 130aa, >Zm00062aa006531_T00 *	
	<    X Cluster 60007 ^ V	>Cluster 60013 0 130aa, >Zm00062aa007889_T00 * Tout surligner Respecter la casse Respecter les accents et diacritiques Mots entiers Occurrence 1 sur 1	I

Compare n gene/protein dataset

From n annotation versions From n individuals (same species)

Input : outputs from UC1

Step 1 files concatenation

Step 2 cluster seq with CD-Hit

Zm00064 : Gaspe Zm00063 : EA1197 Zm00062 : F331

```
>Cluster 60004
        130aa, >Zm00062aa000798 T00... *
0
>Cluster 60005
0
        130aa, >Zm00062aa001058 T00... *
1
        130aa, >Zm00064aa001104_T00... at 100.00%
>Cluster 60006
        130aa, >Zm00062aa002134_T00... *
0
>Cluster 60007
        130aa, >Zm00062aa004172_T00... *
0
        130aa, >Zm00064aa004266 T00... at 100.00%
>Cluster 60008
        130aa, >Zm00062aa004378 T00... *
0
        130aa, >Zm00064aa004472 T00... at 100.00%
1
```

<b>=</b> Galaxy France	🏠 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🕇 📻 📢 🏢
Tools Image: Constraint of the sections   filter fasta Image: Constraint of the sections   Image: Constraint of the sections	<ul> <li>✓ Filter FASTA on the headers and/or the sequences (Galaxy Version 2.3)</li> <li>☆ ▼ ▶ Run Tool</li> <li>Tool Parameters</li> <li>FASTA sequences *</li> <li>1: F331.Gaspe.EA1197.rep.fasta</li> <li>↓ ▷</li> </ul>
sequences Filter fasta to remove sequences based on input criteria (filter_fasta) WORKFLOWS	Criteria for filtering on the headers <ul> <li>Regular expression on the headers</li> <li>Regular expression pattern the header should match *</li> </ul>
All workflows	Zm00064aa004266         Use the Python regular expression syntax as specified in https://docs.python.org/3/library/re.html         Criteria for filtering on the sequences
	No filtering     Remove duplicate sequences     No

### **=** Galaxy France

Tools	☆	•
miniprot	*	×
1 Upload Data		
Show Sections		
<b>Miniprot index</b> build a genome in miniprot	idex f	or

**Miniprot align** align a protein sequence against a genome with affine gap penalty, splicing and frameshift

WORKFLOWS

All workflows

🗲 Miniprot align	☆	&	-	
lign a protein sequence against a genome with affine gap penalty, splicing and frameshift				
Galaxy Version 0.13+galaxy0)				
Tool Parameters				
Database type				
Pre-indexed				
Pre-indexed				
Build an index from FASTA or use a pre-indexed database				
Pre-indexed genomic database *				
L         L         4: Zm00064aa004266 (as twobit)			•	1.
A pre-indexed database built by miniprot				
otein sequence (FASTA) *				
1     1 <td></td> <td></td> <td>•</td> <td>1.</td>			•	1.
otein sequences to be aligned in FASTA format				
utput format *				
GFF3				

🖌 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🕇 📻 📢 🏢

<b>=</b> Galaxy France	🖀 Workflow Visualize Données partagées - Aide - Utilisateur -	≈ ๙
Tools 🔂 🔻	compa ×	pol
miniprot × ×	aligi (Gal Comparative genomics/Gaspe.genome.fasta.gz -	
1 Upload Data	Comparative genomics/F331.rep.fasta -	· ·
Show Sections	Comparative genomics/F331.genome.fasta.gz -	
Miniprot index build a genome index for	Dat Comparative genomics/F331.Gaspe.EA1197.rep.fasta -	
miniprot	P Comparative genomics/EA1197Sub.fasta -	-
Miniprot align align a protein sequence	Bui Comparative genomics/EA1197.rep.fasta -	-
against a genome with affine gap penalty, splicing and frameshift	Pre Comparative genomics/EA1197int.bed -	
WORKFLOWS	C /Comparative genomics/EA1197.genome.mpi -	-
All workflows	A p	-
	Protein sequences to be aligned in FASTA format Dutput format * GFF3	Cancel

#### Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🕆 🚖 📢 🏢

Tools miniprot 1 Upload Dat

Show Section

Miniprot index build a geno miniprot

Miniprot align align a protei against a genome with affine splicing and frameshift

#### WORKFLOWS

All workflows

5							
للا ا		🖋 Miniprot align	☆	&	•	🕨 Rur	n Tool
*	×	align a protein sequence against a genome with affine gap penalty, splicing and frameshift (Galaxy Version 0.13+galaxy0)					
-							
ta		Tool Parameters					
ns		Detekses two					
ome index	for	Database type					
		Pre-indexed					•
in sequen	се	Build an index from FASTA or use a pre-indexed database					)
e gap pena	alty,	Pre-indexed genomic database *					
		D         D         Selected: /Comparative genomics/EA1197.genome.mpi			•	1	
		A pre-indexed database built by miniprot					
		Protein sequence (FASTA) *					
		□     □     4: Zm00064aa004266			•	1	
		Protein sequences to be aligned in FASTA format					
		Output format *					
		GFF3					•

## 🖀 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 🞓 📢 🏢

Seqid	Source	Туре	Start	End	Score	Strand	Phase	Attributes
##gff-v	ersion 3							
##PAF	Zm00064aa004266_T001	130	0	130	-	chr8	193330780	31199353 31199915 366 390 0 AS:i:536 ms:i:616 np:i:124 fs:i:0 st:i:0 da:i:0 do:i:0 cg:Z:44M83U53M92N32M cs:Z::14*tacC:29*gG~gt80ag-gc:2*ggcS:35*aagT:2*gagł 8*cgcC*cagK~gt92ag:18*cgcS:13
chr8	miniprot	mRNA	31199351	31199915	616	-		ID=MP000001;Rank=1;Identity=0.9385;Positive=0.9538;Target=Zm00064aa0042 1 130
chr8	miniprot	CDS	31199783	31199915	210	-	0	Parent=MP000001;Rank=1;Identity=0.9778;Target=Zm00064aa004266_T00114
chr8	miniprot	CDS	31199542	31199702	245	-	2	Parent=MP000001;Rank=1;Identity=0.8868;Target=Zm00064aa004266_T001 45
chr8	miniprot	CDS	31199351	31199449	161	-	0	Parent=MP000001;Rank=1;Identity=0.9688;Target=Zm00064aa004266_T001 99
chr8	miniprot	stop_codon	31199351	31199353	0	-	0	Parent=MP000001;Rank=1

## 🖀 Workflow Visualize Données partagées 🗸 Aide 🗸 Utilisateur 🛪 🞓 📢 🏢

Seqid	Source	Туре	Start	End	Score	Strand	Phase	Attributes
##gff-v	ersion 3							
##PAF	Zm00064aa004266_T001	130	0	130	-	chr8	193330780	31199353 31199915 366 390 0 AS:i:536 ms:i:616 np:i:124 fs:i:0 st:i:0 da:i:0 do:i:0 cg:Z:44M83U53M92N32M cs:Z::14*tacC:29*gG~gt80ag-gc:2*ggcS:35*aagT:2*gagł 8*cgcC*cagK~gt92ag:18*cgcS:13
chr8	miniprot	mRNA	31199351	31199915	616	-		ID=MP000001;Rank=1;Identity=0.9385;Positive=0.9538;Target=Zm00064aa0042 1 130
chr8	miniprot	CDS	31199783	31199915	210	-	0	Parent=MP000001;Rank=1;Identity=0.9778;Target=Zm00064aa004266_T001144
chr8	miniprot	CDS	31199542	31199702	245	-	2	Parent=MP000001;Rank=1;Identity=0.8868;Target=Zm00064aa004266_T001 45
chr8	miniprot	CDS	31199351	31199449	161	-	0	Parent=MP000001;Rank=1;Ideptity=0.9688;Target=Zm00064aa004266_T001 99
chr8	miniprot	stop_codon	31199351	31199353	0	-	0	Parent=MP000001;Rank-1

#### **CD-HIT Use case 3 : extand previous use-case to more than 2 sequences sets**

 >Cluster 60007

 0
 130aa,

 1
 130aa,

 >Zm00064aa004266\_T00...

Compare n gene/protein dataset

#### From n annotation versions From n individuals (same species)

≡		Gas	spe,EA1197		_ ×
≔ : 1:215,356,1	75215,574,168 Q	217,993 bp chr1:221,452,845221,580	, <b>013 Q</b> 127,169 bp		
S	215,400,000	215,440,000 0064aa004265	215,480,000	215,520,000 Zm00064aa004266 Zm00064aa004267	215,560 ~ Q Q Z
chr1 221,460,000	221,480,000 <b>₩</b> ₽ Zm0	221,500,000 0063aa028801	221,520,000	221,540,000 ←(-+) Zm00063aa028802	221,560,000

Workflow	Visualize	Données partagées -	Aide 🔻	Utilisateur 🔻		
	FIGUULEC	Donneed partageed	Aluc	ornibuteur		

Tools	☆	•
getfasta	*	×

🤹 Upload Data

Show Sections

**bedtools getfasta** use intervals to extract sequences from a FASTA file

WORKFLOWS

All workflows

ø beutoots geti	asta use intervals to extract sequences from a FASTA file (Galaxy Version 2.31.1+galaxy0)	☆ 👶	- P	Run
Tool Paran	neters			
ED/bedGraph/GF	F/VCF/EncodePeak file *			
00	Selected: /Comparative genomics/gaspeInt.bed		•	<b>1</b>
bed)				
Choose the sour	ce for the FASTA file			
History				
FASTA file *				
	Selected: /Comparative genomics/Gaspe.genome.fasta.gz			t.
(-fi)				
	lumn in the BED file and the coordinates for the FASTA headers in the output FASTA file			
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No No name) Se the 'name' co	lumn in the BED file and the coordinates for the FASTA headers in the output FASTA file			
se the 'name' col No name) se the 'name' col No nameOnly)	lumn in the BED file and the coordinates for the FASTA headers in the output FASTA file lumn in the BED file for the FASTA headers in the output FASTA file			
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Se the 'name' color No name) Se the 'name' color No nameOnly) eport extract sec	lumn in the BED file and the coordinates for the FASTA headers in the output FASTA file lumn in the BED file for the FASTA headers in the output FASTA file			
Se the 'name' color No name) Se the 'name' color No nameOnly) Seport extract second	lumn in the BED file and the coordinates for the FASTA headers in the output FASTA file lumn in the BED file for the FASTA headers in the output FASTA file quences in a tab-delimited format instead of in FASTA format			

# Tools ☆ ▼ getfasta × × Lupload Data ✓ ✓ ③ Show Sections ✓ ✓

**bedtools getfasta** use intervals to extract sequences from a FASTA file

WORKFLOWS

All workflows

& hadtools gotf:	<b>sta</b> use intervals to extract sequences f	rom a EASTA	filo (Galavi	Version 2.211+gala	20		\$		-
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	Selected: /Comparative genomics/EA11	197int.bed							-
bed)									
Choose the sourc	e for the FASTA file								
History									
FASTA file *									
	Selected: /Comparative genomics/EA	1197.genome.	fasta.gz						- 1
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se the 'name' colu No name) se the 'name' colu No nameOnly)		aders in the c	output FA	STA file	ASTA file				
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# **= Galaxy France** Aide • Utilisateur •

Tools		☆	•
chrom		*	×
	1 Upload Data		
	Show Sections		

**Chromeister** ultra-fast pairwise genome comparisons

**SnpEff chromosome-info:** list chromosome names/lengths

xcms findChromPeaks (xcmsSet) Chromatographic peak detection

**xcms plot chromatogram** Plots base peak intensity chromatogram (BPI) and total ion current chromatogram (TIC) from MSnbase or xcms experiment(s)

xcms findChromPeaks Merger Merge xcms findChromPeaks RData into a unique file to be used by group

xcms groupChromPeaks (group) Perform the correspondence, the grouping of chromatographic peaks within and between samples.

xcms refineChromPeaks (refine) Remove or merge chromatographic peaks based on specific criteria.

*Chromeister* ultra-fast pairwise genome comparisons (Galaxy Version 1.5.a+galaxy1) **Tool Parameters Query sequence** \* C பு 12: EA1197\_cluster60007 Query sequence file in fasta format Reference sequence \* C பு  $\Box$ 9: gaspe\_cluster60007 Reference sequence file in fasta format Output dotplot size \*  $\hat{\phantom{a}}$ 500 Use around 1000 for chromosome-sized sequences and around 2000 for complete genomes K-mer seed size \* 32 Use 32 as default, and 16 in case no similarities are found Diffuse value \*  $\hat{\phantom{a}}$ 1 Level of the heuristic subsampling employed. Change to 1 or 2 if no similarity is found Add grid to plot for multi-fasta data sets

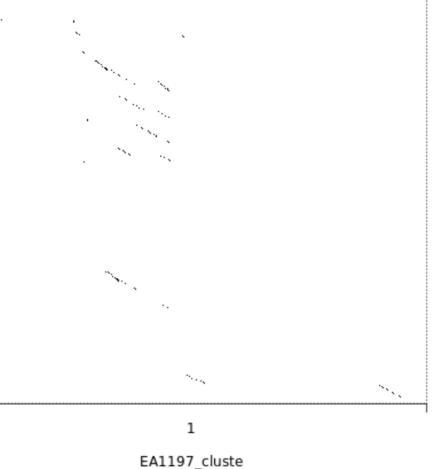
xcms fillChromPeaks (fillPeaks) Integrate

Yes

**=** Galaxy France Workflow Visualize Données partagées - Aide - Utilisateur -Tools ☆ EA1197\_cluster60007-gaspe\_cluster60007.mat filt. score= 0. \* chrom X **1** Upload Data Show Sections Chromeister ultra-fast pairwise genome comparisons gaspe\_cluste SnpEff chromosome-info: list chromosome names/lengths 1 xcms findChromPeaks (xcmsSet) Chromatographic peak detection xcms plot chromatogram Plots base peak intensity chromatogram (BPI) and total ion current chromatogram (TIC) from MSnbase or xcms experiment(s)

xcms findChromPeaks Merger Merge xcms findChromPeaks RData into a unique file to be used by group

xcms groupChromPeaks (group) Perform

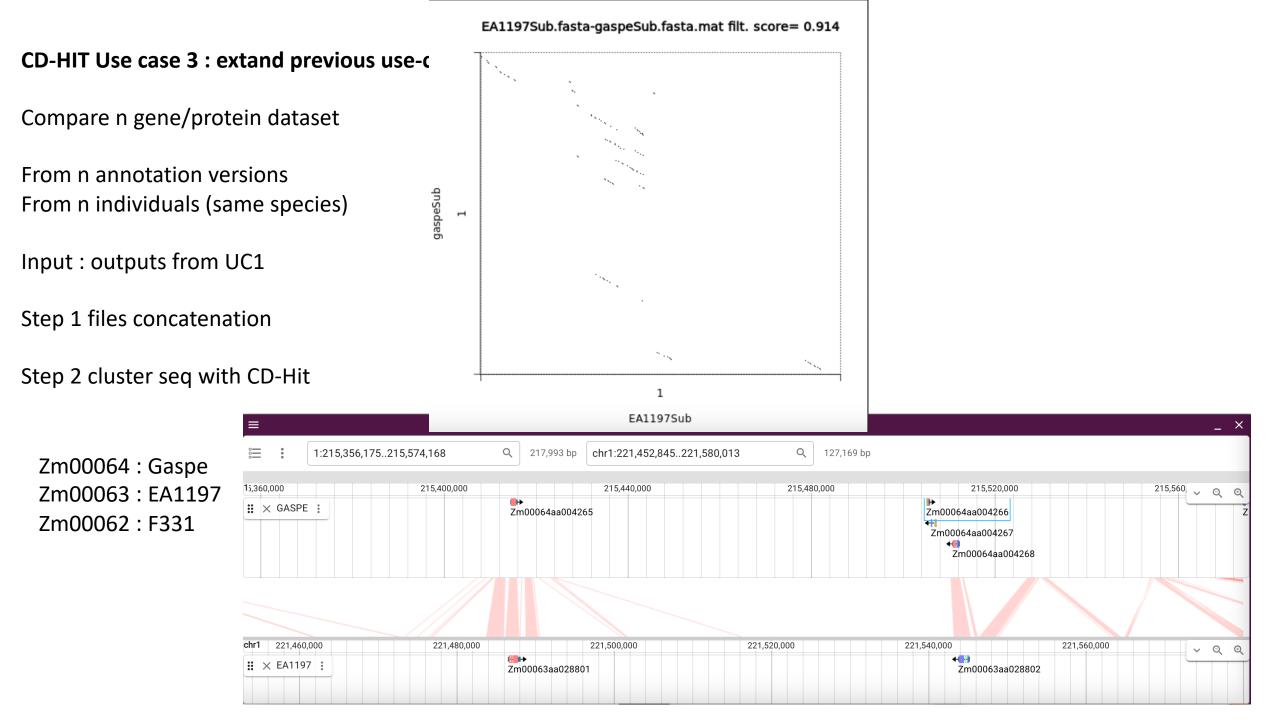


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Tools ☆ •	Chromeister ultra-fast pairwise	compa			🟠 💌 🕨 Run Tool
chrom 🛛 🛠 🗙	Tool Parameters	Label	Details	🛊 Time	
1 Upload Data	Query sequence *	Comparative genomics/gaspeSub.fasta	-	-	
Show Sections	1 (1) (1) (1) (1) (1) (1) (1) (1) (1) (1	Comparative genomics/Gaspe.rep.fasta	-	-	- 1
Chromeister ultra-fast pairwise genome	Query sequence file in fasta format	Comparative genomics/gaspeInt.bed	-	-	
comparisons	Reference sequence *	Comparative genomics/Gaspe.genome.fasta.gz	-	-	
SnpEff chromosome-info: list chromosome names/lengths	1 [] [] [] 4: Zm00064aa	Comparative genomics/F331.rep.fasta	-	-	• <b>1</b> 🕞
xcms findChromPeaks (xcmsSet)		Comparative genomics/F331.genome.fasta.gz	-	-	
Chromatographic peak detection	Reference sequence file in fasta forr Output dotplot size *	Comparative genomics/F331.Gaspe.EA1197.rep.fasta	-	-	
xcms plot chromatogram Plots base peak intensity chromatogram (BPI) and total ion		Comparative genomics/EA1197Sub.fasta	-	-	
current chromatogram (TIC) from MSnbase	1000	Comparative genomics/EA1197.rep.fasta	-	-	
or xcms experiment(s)	Use around 1000 for chromosome-s K-mer seed size *	Comparative genomics/EA1197int.bed	-	-	
xcms findChromPeaks Merger Merge xcms findChromPeaks RData into a unique	K-mer seed size	Comparative genomics/EA1197.genome.mpi	-	-	
file to be used by group	32	<sup>C</sup> /Comparative genomics/FΔ1197 genome fasta gz		-	<b>~</b>
xcms groupChromPeaks (group) Perform	Use 32 as default, and 16 in case no	Back      Lpload		Cancel	
the correspondence, the grouping of chromatographic peaks within and between	Diffuse value *				
samples.	4				•
xcms refineChromPeaks (refine) Remove or merge chromatographic peaks based on	Level of the heuristic subsampling er	nployed. Change to 1 or 2 if no similarity is found			
specific criteria.	Add grid to plot for multi-fasta da	ta sets			
xcms fillChromPeaks (fillPeaks) Integrate	Yes				
areas of missing peaks	Do not use grid if your multi-fasta co	ntains more than a hundred sequences (approximately)			



#### 🖀 Workflow Visualize Données partagées 🖣 Aide 🖣 Utilis

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1 Upload Data	
Show Sections	
Chromeister ultra-fast pairwise geno	ome
comparisons	
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or rems experiment(s)	
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file to be used by group	
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the correspondence, the grouping of	
chromatographic peaks within and be	
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or merge chromatographic peaks bas	sed on
specific criteria.	
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≡			Gaspe,EA1197			
	8:25,283,64125,311,569	Q 27,929 bp	chr8:26,512,74826,5	G37,620 Q	24,872 bp	
884,000	25,288,000	25,292,000	25,296,000 Zm00064aa034370	25,300,000	25,304,000 25 ► Zm00064aa034371	,308,000
chr8	26,516,000	26,520,000	26,524,000	26,528,000	26,532,000	26.5
4	Ze, 516,000 7 1 20063aa038377	26,520,000	Zm00063aa038378	20,528,000	Zm00063aa038379	26,5 Zm0006

Tools	3 -	>Cluster 608
		0 1394aa, >Zm00063aa013167_T00 at 98.78%
chrom 😽	×	1 1396aa, >Zm00062aa018836_T00 *
		2 1396aa, >Zm00064aa019116_T00 at 99.14%
A		>Cluster 609
1. Upload Data		0 1396aa, >Zm00064aa026404_T00 *
		>Cluster 610
Show Sections		0 1396aa, >Zm00064aa040375_T00 * >Cluster 611
Churches where fact a simulate some		0 1392aa, >Zm00063aa036892 T00 at 98.20%
Chromeister ultra-fast pairwise genor	ne	1 1262aa, >Zm00062aa042738_T00 at 98.81%
comparisons		2 1396aa, >Zm00064aa043042_T00 *
SnpEff chromosome-info: list chromo	osome	Cluster C12
	0301110	0 1395aa, > <mark>Zm00063aa038378</mark> _T00 *
names/lengths		1 1395aa, >Zm00062aa034030_T00 at 99.93%
xcms findChromPeaks (xcmsSet)		>Cluster 613
Chromatographic peak detection		0 1395aa, >Zm00062aa021699_T00 *
en en alogiapine peak detection		>Cluster 614 0 1394aa. >Zm00063aa020662 T00 *
xcms plot chromatogram Plots base	peak	0 1394aa, >Zm00063aa020662_T00 * 1 1394aa, >Zm00062aa007996 T00 at 99.93%
intensity chromatogram (BPI) and tota	al ion	2 _ 1394aa, >Zm00064aa008223_T00 at 99.93%
current chromatogram (TIC) from MSn	base	
or xcms experiment(s)		2 1308aa, >21100004aa012937_100 at 99.85
or xems experiment(s)		>Cluster 660
xcms findChromPeaks Merger Merg	е	0 1368aa, > <mark>Zm00064aa034370</mark> _T00 *
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file to be used by group		0 1367aa, >Zm00062aa038519_T00 *
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Acting groupernorm eaks (group) i en	rform	2 1392aa, >Zm00064aa020356_T00 at 99.93%
the company dense the mountain of	rform	>Cluster 617
the correspondence, the grouping of		>Cluster 617 0 1392aa, >Zm00063aa031317_T00 *
the correspondence, the grouping of chromatographic peaks within and bet		>Cluster 617 0 1392aa, >Zm00063aa031317_T00 * 1 1234aa, >Zm00062aa006621_T00 at 97.97%
		>Cluster 617 0 1392aa, >Zm00063aa031317_T00 * 1 1234aa, >Zm00062aa006621_T00 at 97.97% >Cluster 618
chromatographic peaks within and bet samples.	tween	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 *</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re	tween move	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1303aa, &gt;Zm00064aa010182_T00 at 90.77%</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re or merge chromatographic peaks base	tween move	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1303aa, &gt;Zm00064aa010182_T00 at 90.77%</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re	tween move	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77%</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re or merge chromatographic peaks base specific criteria.	tween move ed on	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77% &gt;Cluster 619 0 1391aa, &gt;Zm00064aa041061_T00 * &gt;Cluster 620</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re or merge chromatographic peaks base specific criteria. xcms fillChromPeaks (fillPeaks) Int	tween move ed on	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77% &gt;Cluster 619 0 1391aa, &gt;Zm00064aa041061_T00 * &gt;Cluster 620 0 1146aa, &gt;Zm00063aa013849_T00 at 98.17%</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re or merge chromatographic peaks base specific criteria.	tween move ed on	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77% &gt;Cluster 619 0 1391aa, &gt;Zm00064aa041061_T00 * &gt;Cluster 620 0 1146aa, &gt;Zm00063aa013849_T00 at 98.17% 1 1390aa, &gt;Zm00063aa013849_T00 *</pre>
chromatographic peaks within and bet samples. xcms refineChromPeaks (refine) Re or merge chromatographic peaks base specific criteria. xcms fillChromPeaks (fillPeaks) Int areas of missing peaks	tween move ed on cegrate	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00062aa006621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00062aa009876_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77% &gt;Cluster 619 0 1391aa, &gt;Zm00064aa041061_T00 * &gt;Cluster 620 0 1146aa, &gt;Zm00063aa013849_T00 at 98.17% 1 1390aa, &gt;Zm00063aa013849_T00 * 2 1039aa, &gt;Zm00063aa013849_T00 at 98.85%</pre>
<ul> <li>chromatographic peaks within and bet samples.</li> <li>xcms refineChromPeaks (refine) Re or merge chromatographic peaks base specific criteria.</li> <li>xcms fillChromPeaks (fillPeaks) Intareas of missing peaks</li> <li>bcftools color-chrs plugin Color share</li> </ul>	tween move ed on cegrate	<pre>&gt;Cluster 617 0 1392aa, &gt;Zm00063aa031317_T00 * 1 1234aa, &gt;Zm00063aa02621_T00 at 97.97% &gt;Cluster 618 0 1391aa, &gt;Zm00063aa022626_T00 * 1 1303aa, &gt;Zm00063aa022626_T00 at 98.54% 2 1302aa, &gt;Zm00064aa010183_T00 at 99.77% &gt;Cluster 619 0 1391aa, &gt;Zm00064aa041061_T00 * &gt;Cluster 620 0 1146aa, &gt;Zm00063aa013849_T00 at 98.17% 1 1390aa, &gt;Zm00063aa013849_T00 * 2 1039aa, &gt;Zm00063aa013849_T00 at 98.85% &gt;Cluster 621</pre>
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# **OrthoFinder: phylogenetic orthology inference for comparative genomics**

Emms and Kelly *Genome Biology* (2019) 20:238 https://doi.org/10.1186/s13059-019-1832-y

Genome Biology

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#### SOFTWARE

OrthoFinder: phylogenetic orthology inference for comparative genomics

David M. Emms and Steven Kelly\*

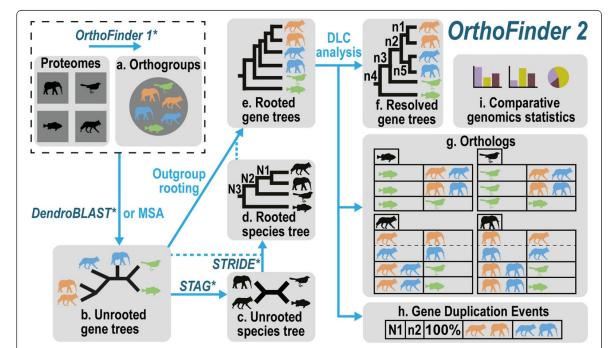


Fig. 2 The OrthoFinder workflow. The method used for each step is shown by the arrow. Published algorithms are shown in italics and are followed by an asterisk. A dotted blue line connecting with a solid arrow indicates additional data that are used in order to carry out the transformation indicated by the solid arrow. MSA, multiple sequence alignment-based tree inference; DLC, duplication-loss-coalescence. (a) Orthogroup inference using the original OrthoFinder algorithm (an orthogroup is the set of genes descended from a single gene in the last common ancestor of all the species under consideration). (b) Gene tree inference. (c) Species tree inference. (d) Species tree rooting (e) Gene tree rooting (f) Hybrid overlap + DLC analysis of rooted gene trees to infer orthologs and gene duplication events. (g) Illustration of the orthologs from the orthologs for each input species (four main boxes). The horizontal divisions within these show the orthologs for each individual species pair. (h) Illustration of the gene duplication event table showing the location of the gene duplication events mapped to the species tree, the percent retention of the duplicate genes in the sampled species, and the genes descended from the gene duplication event. (i) Comparative genomics statistics

## **OrthoFinder best practices : Selecting which species to include**

The first question to ask yourself is what species should you include. The answer to this probably depends on what kind of analysis you want to perform.

Three standard analyses are:

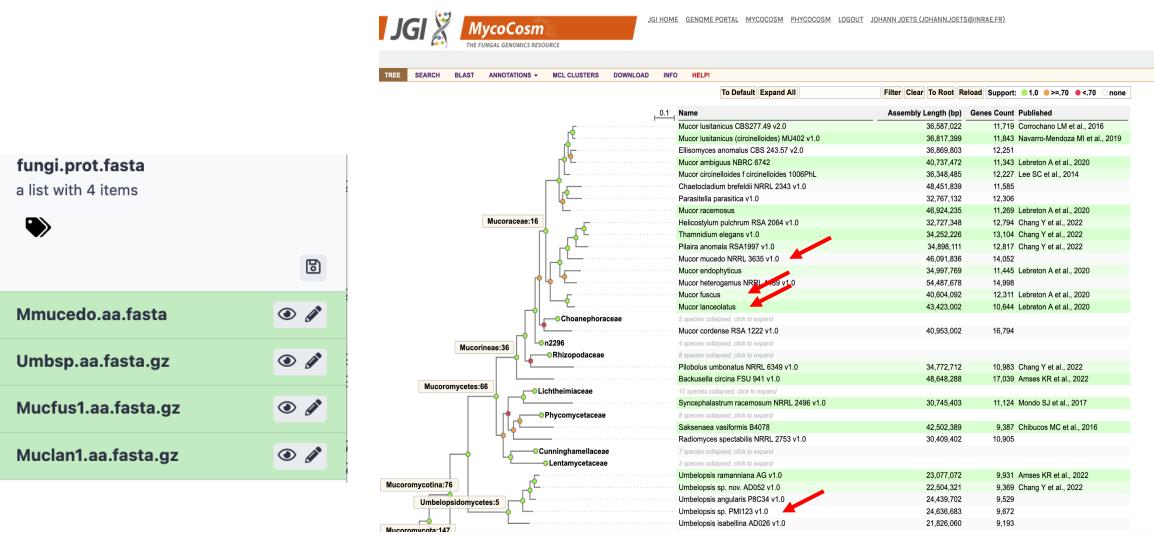
- •Performing a comparative analysis across a clade of species
- •Identifying orthologs between a pair, or among a small number, of species
- •Investigating changes at a particular point in evolutionary history

In the first case just get the proteomes for all the species in your clade that you can. Generally, you don't need to include an outgroup for your clade of interest—in fact, this will push back the point in evolutionary history at which your orthogroups are defined (<u>Orthogroups, Orthologs & Paralogs</u>) and so it's usually better not to since your orthogroups will have lower resolution.

In the second case, it is good to ensure you have sufficient species sampling so as to get the best results. The same rule applies as for inferring a good phylogenetic tree: you should break up long branches with intermediate species. You want an absolute minimum of 4 species and somewhere between 6-10 is probably optimal.

If you're interested in what happened on a particular branch of the species tree, then you should likewise ensure good species sampling—ideally at least two species below the branch, at least two species on the closest branch above and two or more species in the outgroup.

# **OrthoFinder. Our dataset:**



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UseGalaxy.fr will be undergoing maintenance from	October 6th to 7th. Running jobs will be stopped. Thank you for your understanding.					
Tools ☆ ≔	☆ IE Executed OrthoFinder and successfully added 1 job to the queue.					
orthofinder 😢	The tool uses this input:					
1 Upload Data	• 10: fungi.prot.fasta (with implicit datatype conversion)					
Show Sections	It produces 12 outputs:					
<b>OrthoFinder</b> finds orthogroups in a set of proteomes	<ul> <li>20: OrthoFinder on data 5, data 6, and others: orthogroups (txt)</li> <li>21: OrthoFinder on data 5, data 6, and others: orthogroups (tsv)</li> <li>22: OrthoFinder on data 5, data 6, and others: species overlaps</li> <li>23: OrthoFinder on data 5, data 6, and others: unassigned genes</li> </ul>					
WORKFLOWS						
All workflows	<ul> <li>24: OrthoFinder on data 5, data 6, and others: overall comparative genomics statistics</li> <li>25: OrthoFinder on data 5, data 6, and others: per species comparative genomics statistics</li> <li>26: OrthoFinder on data 5, data 6, and others: species tree</li> <li>27: OrthoFinder on data 5, data 6, and others: species tree with node labels</li> <li>28: OrthoFinder on data 5, data 6, and others: species tree with duplication events</li> <li>29: OrthoFinder on data 5, data 6, and others: duplication events</li> <li>30: OrthoFinder on data 5, data 6, and others: duplications per orthogroup</li> <li>31: OrthoFinder on data 5, data 6, and others: duplications per species tree node</li> </ul> You can check the status of queued jobs and view the resulting data by refreshing the History panel. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.					

• / × 27: OrthoFinder on data 5, data 6, and others: spe cies tree with node labels 1 line format: newick, génome de référence: ? OrthoFinder version 2.5.4 Copyright (C) 2014 David Emms 2022-09-28 04:21:27 : Starting OrthoFinder 2.5.4 1 thread(s) for highly parallel tasks (BLAST searches etc.) 1 thread(s) for OrthoFinder algorithm Checking required programs are installed ₿**₽**₿С!!!!? 

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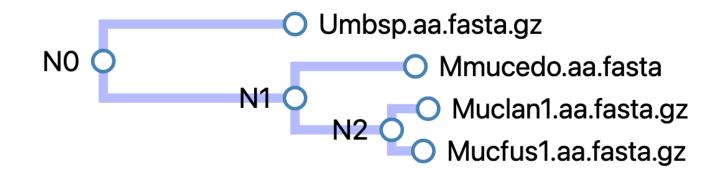
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Tools ☆	E search visualizations
search tools	
1 Upload Data	Editor Manually edit text
Get Data	
Send Data	Phylogenetic Tree Visualization     A performant, reusable, and extensible tree visualisation library for the web hosted at: http://biojs.io/d/phylocanvas.
<b>Collection Operations</b>	
GENERAL TEXT TOOLS	Phyloviz
Text Manipulation	Phylogenetic data analysis from multiple data sources.
Filter and Sort	
Join, Subtract and Group	
GENOMIC FILE MANIPULATION	
Convert Formats	
FASTA/FASTO	

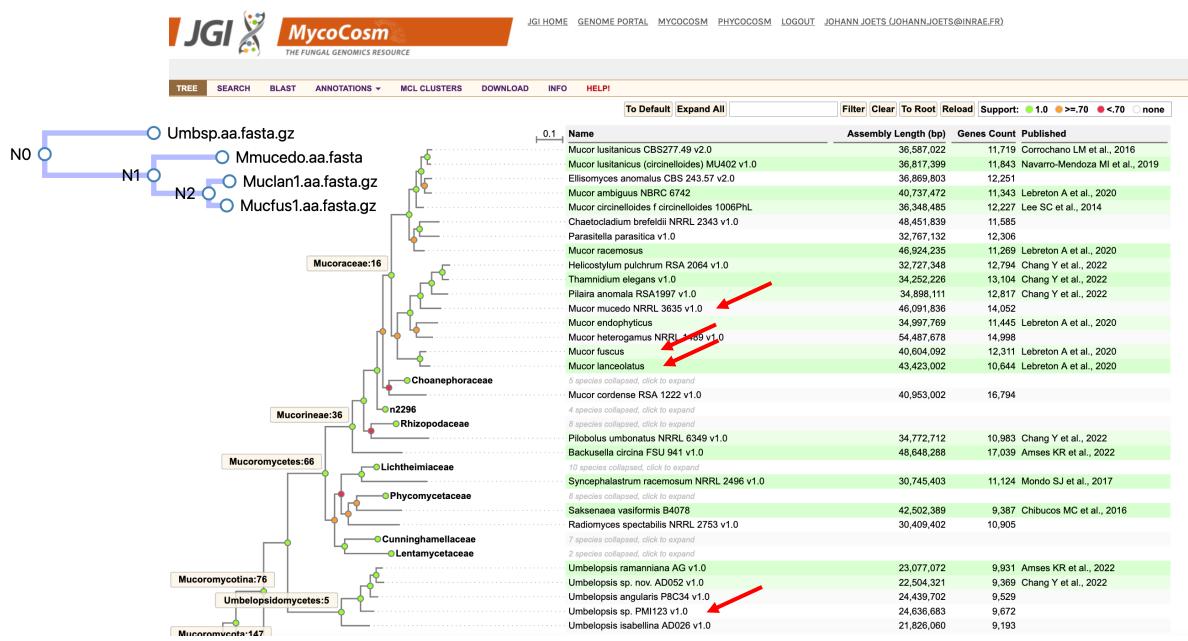
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Get Data	
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<b>Collection Operations</b>	
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Text Manipulation	Phylogenetic data analysis from multiple data sources.
Filter and Sort	
Join, Subtract and Group	
GENOMIC FILE MANIPULATION	
Convert Formats	
FASTA/FASTO	

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Phylogenetic Tree from OrthoFinder on data 5, data 6, and others: species tree with node labeis: | Alt+click to select nodes





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Number of species	4
Number of species	4
Number of genes	45727
Number of genes in orthogroups	42977
Number of unassigned genes	2750
Percentage of genes in orthogroups	94.0
Percentage of unassigned genes	6.0
Number of orthogroups	9227
Number of species-specific orthogroups	707
Number of genes in species-specific orthogroups	3857
Percentage of genes in species-specific orthogroups	8.4
Mean orthogroup size	4.7
Median orthogroup size	4.0
G50 (assigned genes)	4
G50 (all genes)	4
O50 (assigned genes)	2851
O50 (all genes)	3195
Number of orthogroups with all species present	5270
Number of single-copy orthogroups	3448
Date	2022-09-28
Orthogroups file	Orthogroups.tsv
Unassigned genes file	Orthogroups_UnassignedGenes.tsv
Per-species statistics	Statistics_PerSpecies.tsv
Overall statistics	Statistics_Overall.tsv
Orthogroups shared between species	Orthogroups_SpeciesOverlaps.tsv
Average number of genes per-species in orthogroup	Number of orthogroups Percentage of orthogroups Number of genes Percentage of genes
<1	3049 33.0 8180 19.0
'1	5328 57.7 24299 56.5
'2	573 6.2 5160 12.0
'3	141 1.5 1855 4.3
'4	63 0.7 1089 2.5
'5	20 0.2 428 1.0

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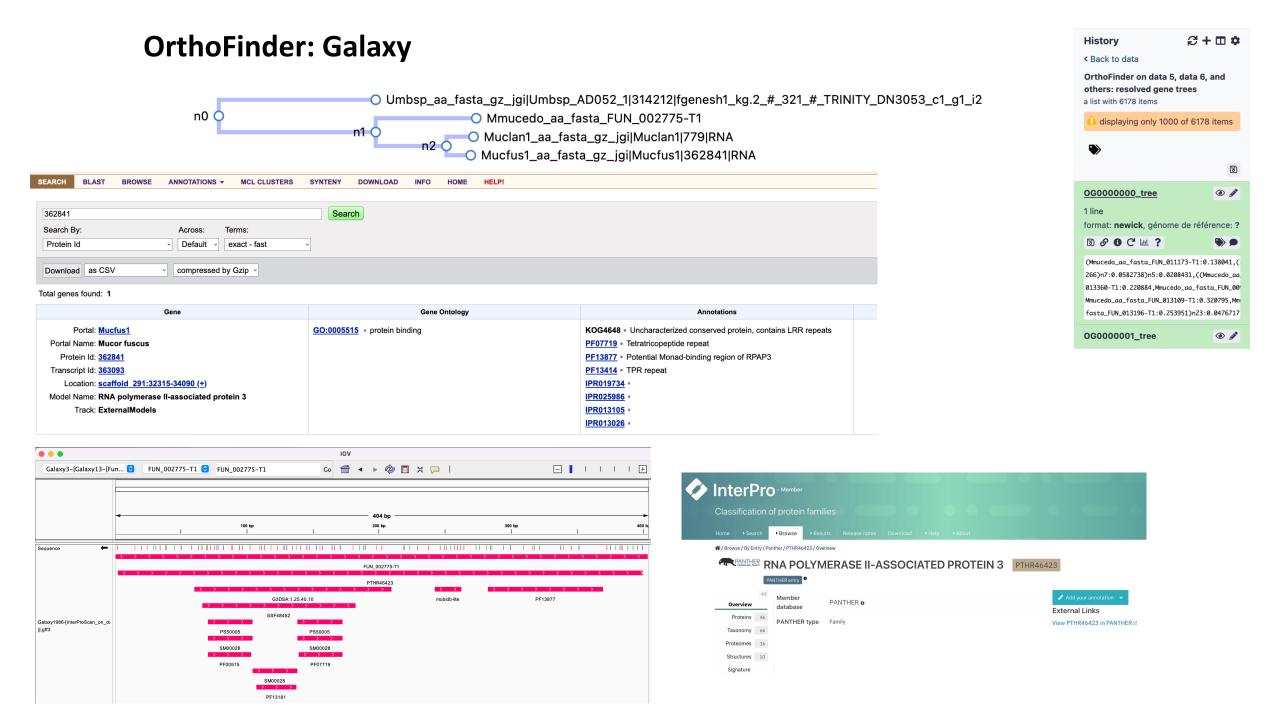
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Number of genes	13403	12311	10644	9369	Rechercher des donnees
Number of genes in orthogroups	12626	11987	10268	8096	data
Number of unassigned genes	777	324	376	1273	30 shown, 1 deleted, 6202 hidden
Percentage of genes in orthogroups	94.2	97.4	96.5	86.4	
Percentage of unassigned genes	5.8	2.6	3.5	13.6	352.53 MB
Number of orthogroups containing species	7814	8339	8153	6236	
Percentage of orthogroups containing species	84.7	90.4	88.4	67.6	25: OrthoFinder on data 💿 🖋 🗙
Number of species-specific orthogroups	330	130	58	189	5, data 6, and others: pe
Number of genes in species-specific orthogroups	2454	567	198	638	r species comparative genomics stati stics
Percentage of genes in species-specific orthogroups	18.3	4.6	1.9	6.8	
Number of genes per-species in orthogroup	Number of orthogroups	Number of orthogroups	Number of orthogroups	Number of orthogroups	99 lines, 1 comments
'0	1413	888	1074	2991	format: tsv, génome de référence: ?
'1	6076	6705	6686	5002	OrthoFinder version 2.5.4 Copyright
'2	1119	1110	1114	907	(C) 2014 David Emms
'3	269	241	225	202	2022-09-28 04:21:27 : Starting
'4	97	93	68	66	OrthoFinder 2.5.4
'5	59	55	22	29	1 thread(s) for highly parallel tasks
'6	42	34	19	12	(BLAST searches etc.)
'7	22	12	10	3	1 thread(s) for OrthoFinder algorithm
'8	14	15	2	3	Checking required programs are
'9	18	6	1	1	installed
'10	6	7	2	3	
11-15	35	33	2	5	₿&₿С₩? ♥♥
16-20	24	10	1	3	
21-50	27	17	1	0	1. 2.Mmuce
51-100	5	1	0	0	Mmucedo
101-150	1	0	0	0	Number of genes13403Number of genes in orthogroups12626
151-200	0	0	0	0	Number of unassigned genes 777
201-500	0	0	0	0	Percentage of genes in orthogroups 94.2
501-1000	0	0	0	0	
'1001+	0	0	0	0	24: OrthoFinder on data 💿 🖉 🗙
Number of genes per-species in orthogroup	Percentage of orthogroups	Percentage of orthogroups	Percentage of orthogroups	Percentage of orthogroups	5, data 6, and others: ove rall comparative genomics statistics
'0	15.3	9.6	11.6	32.4	ran comparative genomics statistics
'1	65.9	72.7	72.5	54.2	23: OrthoFinder on data 💿 🖋 🗙
'2	12.1	12.0	12.1	9.8	5, data 6, and others: una
'3	2.9	2.6	2.4	2.2	ssigned genes
		10	^7	0.7	22. OrthoEindor on data 🖉 🕹 🗙

Number of genes per-species in orthogroup	Percentage of genes	Percentage of genes	Percentage of genes	Percentage of genes	
'0	0.0	0.0	0.0	0.0	
'1	45.3	54.5	62.8	53.4	
'2	16.7	18.0	20.9	19.4	
'3	6.0	5.9	6.3	6.5	- 11
'4	2.9	3.0	2.6	2.8	
'5	2.2	2.2	1.0	1.5	
'6	1.9	1.7	1.1	0.8	
'7	1.1	0.7	0.7	0.2	
'8	0.8	1.0	0.2	0.3	
'9	1.2	0.4	0.1	0.1	
'10	0.4	0.6	0.2	0.3	
11-15	3.2	3.4	0.2	0.7	
16-20	3.2	1.4	0.2	0.5	

25: OrthoFinder on data 5, data 6, and others: pe r species comparative genor stics	● 🧨 🗙 nics stat
99 lines, 1 comments	
format: <b>tsv</b> , génome de référe	nce: ?
OrthoFinder version 2.5.4 Cop	oyright
(C) 2014 David Emms	
2022-09-28 04:21:27 : Starti	ng
OrthoFinder 2.5.4	
1 thread(s) for highly parallel	tasks
(BLAST searches etc.)	
1 thread(s) for OrthoFinder al	gorithm

	•			5, data 6, and others: ort
			Q~ Rec	hogroups (tsv)
●●● 合品 ち* び =	Classeur3		Q~ Rec	5 1 1 1 1
Accueil Insertion Dessin Mise en page	Formules Données Révision Affichage			9,227 lines, 1 comments
Calibri (Corps) V 12 V A^ AV	Image: standard     Image: standard	r Insérer ▼	∑ × As ▼ Z	format: <b>tsv</b> , génome de référence: <b>?</b>
		Mise en forme Mettre sous Styles de conditionnelle forme de tableau cellule Mise en form		OrthoFinder version 2.5.4 Copyright
Ouvrir des classeurs récupérés ? Vos modifications ré	centes ont été enregistrées. Voulez-vous continuer à travailler là où vou	us vous étiez arrêté ?		(C) 2014 David Emms
A6042 🔹 🗙 🗸 🏂 OG0006040				
АВ	C	D		2022-09-28 04:21:27 : Starting
5818 0G0005816 FUN_011186-11	Jgi   Muctus 1   369267   Mt 11816.m_SAM50-IIKe	Jgi   Mucian1   1369   SAM50-IIKe	Jgi   Umbsp_ADU52	
5819 OG0005817 FUN_011187-T1	jgi   Mucfus1   358331   Mf86363.m_Unknown	jgi   Muclan1   2915   Ml92955.m_Unknown	jgi Umbsp_AD052	OrthoFinder 2.5.4
5820 OG0005818 FUN_011190-T1	jgi   Mucfus1   358334   Guanosine-diphosphatase	jgi   Muclan1   2918   Guanosine-diphosphatase	jgi   Umbsp_AD052	(three d(a) for bighty negative to de-
5821 OG0005819 FUN_011197-T1	jgi   Mucfus1   357467   Pirin	jgi Muclan1 3538 Pirin	jgi Umbsp_AD052	1 thread(s) for highly parallel tasks
5822 OG0005820 FUN 011202-T1	jgi   Mucfus1   357463   Signal	jgi   Muclan1   3534   Signal	jgi Umbsp_AD052	(BLAST searches etc.)
5823 OG0005821 FUN 011209-T1	jgi Mucfus1 358899 Bloom, jgi Mucfus1 360777 ATP-depende		JB. 1 011103P_1 (20032	
5824 OG0005822 FUN_011213-T1	jgi Mucfus1 368705 Mf96070.m_SH3	jgi Muclan1 7256 Ml07161.m_SH3	jgi Umbsp_AD052	1 thread(s) for OrthoFinder algorithm
5825 OG0005823 FUN_011216-T1	jgi Mucfus1 358055 Gamma-secretase	jgi Muclan1 2952 Gamma-secretase	jgi Umbsp_AD052	
5826 OG0005824 FUN_011221-T1	jgi   Mucfus1   365074   Mf10441.m_Unknown	jgi Muclan1 7773 MI56041.m_Unknown	jgi Umbsp_AD052	Chapking required programs are
5827 OG0005825 FUN_011222-T1	jgi Mucfus1 366442 Multiprotein-bridging	jgi Muclan1 9299 Multiprotein-bridging	jgi Umbsp_AD052	Checking required programs are
5828 OG0005826 FUN_011258-T1	jgi Mucfus1 357049 Mf96233.m_Unknown	jgi Muclan1 997 Ml64269.m_Unknown	jgi Umbsp_AD052	installed
5829 OG0005827 FUN_011314-T1	jgi Mucfus1 359380 Mf62724.m_Unknown	jgi Muclan1 3091 Ml44294.m_Unknown	jgi Umbsp_AD052	motuneu
5830 OG0005828 FUN_011315-T1	jgi Mucfus1 359378 Poly_rCbinding	jgi Muclan1 3090 Poly_rCbinding	jgi Umbsp_AD052	
5831 OG0005829 FUN_011316-T1	jgi Mucfus1 366560 Mf17297.m_Unknown	jgi Muclan1 3089 Ml49330.m_Unknown	jgi Umbsp_AD052	
5832 OG0005830 FUN_011318-T1	jgi Mucfus1 359384 L-lactate	jgi Muclan1 1368 L-lactate	jgi Umbsp_AD052	
5833 OG0005831 FUN_011322-T1	jgi   Mucfus1   367884   Uracil-regulated	jgi Muclan1 1627 Uracil-regulated	jgi Umbsp_AD052	ⓑ 𝚱 ϴ Ϲ ···· ?
5834 OG0005832 FUN_011323-T1	jgi   Mucfus1   367885   Uracil	jgi Muclan1 1626 Uracil	jgi Umbsp_AD052	
5835 OG0005833 FUN_011324-T1	jgi Mucfus1 367887 Mf08238.m_Unknown	jgi Muclan1 1625 Ml25463.m_Unknown	jgi Umbsp_AD052	1.Orthogroup 2.Mmucedo.aa.fasta
5836 OG0005834 FUN_011331-T1	jgi Mucfus1 367888 Mf25101.m_ATP-dependent	jgi Muclan1 1621 Ml24741.m_ATP-dependent	jgi Umbsp_AD052	1. or chogroup 2. Milaceuo. au. Tustu
5837 OG0005835 FUN_011344-T1	jgi   Mucfus1   367677   Ubiquitin-conjugating	jgi   Muclan1   4994   Ubiquitin-conjugating	jgi Umbsp_AD052	Orthogroup Mmucedo.aa.fasta
5838 OG0005836 FUN 011346-T1	jgi   Mucfus1   360540   Ribosomal	jgi   Muclan1   10260   Ribosomal	jgi Umbsp_AD052	or energieup mateueraurrasea
5839 OG0005837 FUN_011347-T1	jgi   Mucfus1   357212   Cell	jgi Muclan1 4975 Septin	jgi   Umbsp_AD052	OG0000000 FUN_000055-T1, FUN_000056-T1
	jgi   Mucfus1   359874   Mf15169.m_Unknown	jgi Muclan1 318 Ml56381.m_Unknown	jgi   Umbsp_AD052	
5841 OG0005839 FUN_011354-T1	jgi   Mucfus1   367680   Box	jgi Muclan1 320 Box	jgi   Umbsp_AD052	OG0000001 FUN_000167-T1, FUN_000491-T1
5842 OG0005840 FUN_011356-T1	jgi Mucfus1 367678 Mf26248.m_Unknown	jgi   Muclan1   322   Ml09600.m_Unknown	jgi Umbsp_AD052	0G0000002 FUN_000685-T1. FUN_000919-T1.
5843 OG0005841 FUN 011384-T1	jgi   Mucfus1   365938   Haloacid	jgi   Muclan1   5842   Haloacid	jgi Umbsp_AD052	0G0000002 FUN_000685-T1, FUN_000919-T1
5844 OG0005842 FUN 011386-T1	jgi   Mucfus1   365936   Carbamoyl-phosphate	jgi   Muclan1   5840   Carbamoyl-phosphate	jgi Umbsp_AD052	0G0000003 FUN_002416-T1, FUN_002419-T1
5845 OG0005843 FUN 011390-T1	jgi   Mucfus1   366173   FKBP12-associated	jgi Muclan1 4245 FKBP12-associated	jgi Umbsp_AD052	
5846 OG0005844 FUN_011392-T1	jgi   Mucfus1   367157   Cytoplasmic	jgi   Muclan1   6507   Cytoplasmic		1 312247 estExt_Genewise1Plus.
5847 OG0005845 FUN_011392-11	jgi   Mucfus1   365803   Transcription	jgi Muclan1 7889 Transcription		1 361134 gm1.5473_g
5847 0G0005845 FUN_011394-11 5848 0G0005846 FUN_011396-T1	jgi   Mucfus1   365804   Mf89779.m_Unknown	jgi Muclan1 7899 Ml16212.m_Unknown		1 301134 gm1.5473_g 1 325631 fgenesh1_kg.18_#_70_#
5849 OG0005847 FUN 011442-T1	jgi Mucfus1 359948 NudC	jgi Muclan1 4879 NudC	Igi umbsp AD052	1 313267 fgenesh1 kg.1 # 570 #
Feuil1 +				
Modifier 🎇 Accessibilité : vérification terminée				+ 100 %

#### 21: OrthoFinder on data • / × 5. data 6. and others: ort



October 6th to 7th. Running jobs will be stopped. Thank you for your understanding.

Orthogroup	Mmucedo.aa.fasta	Mucfus1.aa.fasta.gz	Muclan1.aa.fasta.gz	History 🔂 🕂 🖽 🌣
Orthogroup	Mmucedo.aa.fasta	Mucfus1.aa.fasta.gz	Muclan1.aa.fasta.gz	Rechercher des données 🛛 😮 😢
OG0009227	FUN_000003-T1			Rechercher des donniees
OG0009228	FUN_000045-T1			data
OG0009229	FUN_000085-T1			30 shown, 1 deleted, 6202 hidden
OG0009230	FUN_000089-T1			352.53 MB
OG0009231	FUN_000101-T1			352.53 MB
OG0009232	FUN_000124-T1			25: OrthoFinder on data 🔹 🖉 🖈 🗙
OG0009233	FUN_000125-T1			5, data 6, and others: per
OG0009234	FUN_000126-T1			species comparative genomics statis
OG0009235	FUN_000131-T1			ics
OG0009236	FUN_000132-T1			24: OrthoFinder on data 🛛 💿 🧳 🗙
OG0009237	FUN_000133-T1			5, data 6, and others: ove
OG0009238	FUN_000134-T1			rall comparative genomics statistics
OG0009239	FUN_000135-T1			
OG0009240	FUN_000152-T1			23: OrthoFinder on data ④ 🖋 🗙 5, data 6, and others: un
OG0009241	FUN_000157-T1			assigned genes
OG0009242	FUN_000159-T1			2,750 lines, 1 comments
OG0009243	FUN_000176-T1			format: <b>tsv</b> , génome de référence: ?
OG0009244	FUN_000180-T1			
OG0009245	FUN_000187-T1			OrthoFinder version 2.5.4 Copyright
OG0009246	FUN_000243-T1			(C) 2014 David Emms
OG0009247	FUN_000254-T1			2022-09-28 04:21:27 : Starting
OG0009248	FUN_000290-T1			OrthoFinder 2.5.4
OG0009249	FUN_000292-T1			1 thread(s) for highly parallel tasks
OG0009250	FUN_000326-T1			(BLAST searches etc.)
OG0009251	FUN_000337-T1			1 thread(s) for OrthoFinder algorithm
OG0009252	FUN_000340-T1			Checking required programs are
OG0009253	FUN_000450-T1			installed
OG0009254	FUN_000474-T1			
OG0009255	FUN_000498-T1			ਡਿ <b>&amp; 8 ℃</b> ਘ ? 🕨 🔊
060009256	FUN 000515-T1			